

Allogamy, metaxenia and hybrids in orchids, what do we know about it?

[©]José Martín Barreda-Castillo¹, [©]Emerson Ricardo Pansarin², [©]Juan Luis Monribot-Villanueva¹, ¹⁰ José A. Guerrero-Analco¹ and ¹⁰ Rebeca Alicia Menchaca-García 3*

¹Red de Estudios Moleculares Avanzados, Instituto de Ecología (INECOL), A.C., Xalapa, Veracruz, Mexico.

² Departamento de Biologia, Faculdade de Filosofia, Ciências e Letras, Universidade de São Paulo, São Paulo, Brazil.

³Centro de Investigaciones Tropicales (CITRO), Universidad Veracruzana, Xalapa, Veracruz, Mexico.

*Author for correspondence: joseantonio.guerrero@inecol.mx; rmenchaca@uv.mx

Abstract

Orchids are a diverse group of plants, also manifested in their great diversity of flowers. Despite this, orchids are pollinated either through autogamy or allogamy (geitonogamy and xenogamy). Although there are some autogamous orchids, the majority are allogamous, mainly pollinated by xenogamy since they present physical or genetic barriers that prevent self-pollination. In addition, orchids are known for their capacity for interspecific pollination, which could influence fruits (metaxenia) and seeds (xenia) production. Its capacity for hybridization represents an opportunity to produce organisms tolerant to biotic or abiotic stress, in addition to exhibiting new shapes, colors and fragrances; this would be important in horticulture, where the proper selection of parents provides those advantages to the descendant hybrids. This review addresses the characteristics of each type of reproduction systems in orchids, as well as their advantages and disadvantages. At the same time, the study of the induction effect of metaxenia and xenia in this family is proposed. Finally, the production of orchid hybrids is contextualized and the opportunities of this approach in the near future.

Keywords: Autogamy, geitonogamy, genetic improvement, pollination, xenogamy.

Resumen

Las orquídeas son un grupo diverso de plantas, manifestado además en su gran diversidad de flores. A pesar de esto, las orquídeas son polinizadas ya sea mediante autogamia o alogamia (geitonogamia y xenogamia). Si bien existen algunas orquídeas autógamas, la mayoría son alógamas, principalmente polinizadas por xenogamia, ya que presentan barreras físicas o genéticas que evitan la autopolinización. Además, las orquídeas son conocidas por su capacidad de polinización interespecífica, la cual podría influenciar la producción de frutos (metaxenia) y de semillas (xenia). Su capacidad de hibridación representa una oportunidad para producir organismos tolerantes a estrés biótico o abiótico, además de exhibir nuevas formas, colores y fragancias; esto sería importante en horticultura, donde la selección adecuada de los parentales proporciona esas ventajas a los híbridos descendientes. En esta revisión se abordan las características de cada tipo de sistema de reproducción en orquídeas, así como sus ventajas y desventajas. A su vez, se propone el estudio del efecto de inducción de metaxenia y xenia en esta familia. Finalmente, se contextualiza la producción de híbridos de orquídeas, y las oportunidades de este enfoque en el futuro próximo. Palabras clave: Autogamia, geitonogamia, mejoramiento genético, polinización, xenogamia.

This is an open access article distributed under the terms of the Creative Commons Attribution License CCBY-NC (4.0) international. https://creativecommons.org/licenses/by-nc/4.0/

rchidaceae is one of the largest family within the flowering plants (Chase *et al.* 2015, Antonelli *et al.* 2023). This family includes approximately 27,000 species grouped in 750 genera (Lu *et al.* 2019, Antonelli *et al.* 2023). Orchidaceae is subdivided in five subfamilies: Apostasioidae, Vanilloideae, Cypripedioideae, Orchidoideae, and Epidendroideae (Chase *et al.* 2015). The phylogenetic relation among the subfamilies is shown in Figure 1. Orchids are mostly long-lived herbs (Hew & Yong 2004) and are distributed worldwide (except for polar and desert regions), particularly diverse in the tropics (Givnish *et al.* 2015). Most of the orchid species are epiphytes (Rasmussen & Rasmussen 2018), however, some of them are terrestrials, rupicolous or occur in aquatic environments (Shefferson *et al.* 2020).

Orchidaceae is considered as the most evolved plant family due their morpho-physiological particularities (Soltis *et al.* 2019, Hietz *et al.* 2022), such as the presence of pseudobulb and succulent leaves (Ng & Hew 2000), velamentous roots (Zotz & Winkler 2013, Pridgeon 2014), or the presence of crassulacean acid metabolism (Zhang *et al.* 2018). Another interesting feature about the biology of orchids is their seeds. Most orchids produce thousands of dust-like seeds with a non-developed embryo and no endosperm (Yeung 2017). For this reason, orchid seeds need to establish a symbiotic relationship with a mycorrhizal fungus, enabling germination (Otero *et al.* 2004, 2007, Porras-Alfaro & Bayman 2007). Symbiotic interactions between orchid seeds and mycorrhizae are a widely studied topic (Rasmussen 2002, Dearnaley 2007, McCormick *et al.* 2018, Sarsaiya *et al.* 2019, Favre-Godal *et al.* 2020, Li *et al.* 2021b, Selosse *et al.* 2022).

Orchids are remarkable by their flower diversity, much appreciated by the ornamental plant market. Among the ornamental orchids, the genera *Cattleya*, *Cymbidium*, *Dendrobium*, *Phalaenopsis*, *Phaius*, *Paphiopedilum* and *Vanda* are the most cultivated (Vij & Pathak 2012). Even though the flower is the most appreciated part of the orchids, some species such as *Vanilla planifolia* Andrews produce fleshy fruits, commercialized because of their aromatic and flavoring traits (Pérez-Silva *et al.* 2021, da Silva-Oliveira *et al.* 2022). Most orchid species exhibit hermaphrodite flowers, except for some species from the subtribes Catasetinae and Satyriinae, which produce unisexual flowers (Suetsugu 2020), whereas some orchids exhibit flowers whose sexual function is separated in time (Hurskainen *et al.* 2017).

The vast diversity of flowers makes orchids to be highly appreciated among collectors, resulting in overexploitation and trafficking of species. This is the main reason why approximately 1,970 species of orchids are threatened, according to the IUCN red list (Hinsley *et al.* 2018, IUCN 2023). For this reason, Orchidaceae is one of most heavily protected plant family, with comprehensive CITES (Convention on International Trade in Endangered Species of Wild Fauna and Flora) trade restrictions on the entire family; as a result, orchids represent almost 75 % of all CITESlisted species (Phelps 2015).

Several studies on orchids have focused on the diversity of floral characters (Pellegrino *et al.* 2017, Naczk *et al.* 2018, Dellinger 2020, Hu *et al.* 2020), flower rewards and pollinators (Cozzolino & Widmer 2005, Pansarin *et al.* 2008, Pansarin 2016, Fay 2018, Shrestha *et al.* 2020, Castro *et al.* 2021, Ray & Gillett-Kaufman 2022), and aromatic and other chemical compounds (Bohman *et al.* 2016, Wu *et al.* 2019, Ramya *et al.* 2020, Brzosko & Mirski 2021). In fact, several topics on orchid pollination have been studied since Darwin (1862). Here we aim to provide an update on the knowledge about the different types of reproduction systems observed in orchids (autogamy, allogamy, and xenia and metaxenia induction), along with the production of hybrids, with the purpose of promoting both the conservation and improvement of orchids through different pollination strategies.

Pollination. In general, pollination is defined as the transfer of pollen from the anthers to the stigma (Wurz *et al.* 2021). In most of the flowering plants, only a portion of the pollen produced by the anther is carried to the stigma in a single pollination event (Edlund *et al.* 2004). This is also true for the orchid subfamily Apostasioideae (Figure 1), and some Vanilloideae, such as in *Cleistes (e.g.,* Pansarin 2003), in which the pollen is released as free monads. Pollination in Apostasioideae is more reminiscent of that observed in other families than in Orchidaceae (Kocyan & Endress 2001, Yin *et al.* 2016, Li *et al.* 2023). In the most of Orchidaceae subfamilies, the entire anther content

must be removed by the pollinator (Endress 2016). In addition, one or more entire pollen package (*i.e.*, pollinium) are deposited in the stigmatic surface in a single pollination event (*e.g.*, Epidendroideae). As consequence, pollination in orchids has even been considered to be more specialized than other flowering plants (Jersáková *et al.* 2006). However, at the same time it is risky: if the pollinarium is removed by an inefficient pollinator, which can result in pollen lost, fruit set would be compromised (Cabrera-Reyes *et al.* 2021). For this reason, orchids commonly have specialized traits to attract effective pollinators, in order to ensure fruit set (Jersáková *et al.* 2006, Phillips *et al.* 2020).

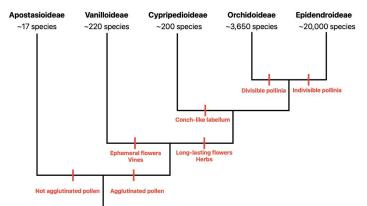


Figure 1. Subfamilies of Orchidaceae. Some differential traits among the subfamilies are highlighted in red. Figure constructed with data from literature (Chase 2005, Yin *et al.* 2016, Hu *et al.* 2022, Jolman *et al.* 2022, dos Santos & da Silva 2023, Kim *et al.* 2023).

In most of the orchids, flowers do not produce any kind of rewards for pollinators, therefore, they are pollinated by food deceit (Pansarin *et al.* 2008), sexual deception or attraction (Cozzolino & Widmer 2005, Mant *et al.* 2005, Shrestha *et al.* 2020, Luo *et al.* 2021). Pollinators commonly land on a flower or on an inflorescence in order to search for a flower reward, commonly nectar or fragrances, which may or may not be present (*i.e.*, deceptive flowers) (Pemberton 2013, Lozano-Rodríguez *et al.* 2022). Besides the production of a flower resource, pollen is deposited on the body of pollinators. Pollen deposition on the stigma occurs when a pollinator carrying pollen visit another flower (Pemberton 2013, Hallett *et al.* 2017). Another important pic in pollination ecology studies is the source of pollen (Tremblay *et al.* 2005, Kropf & Renner 2008), since orchids can reproduce by autogamy, allogamy, or both (Willmer 2011, Bateman 2020). As consequence, in pollination ecology studies it is recommended to carry out pollination by hand, in order to check their reproductive systems, or even to verify whether agamospermy could or could not occur (Tremblay *et al.* 2005, Sao Leao *et al.* 2019, Wurz *et al.* 2021).

Autogamy. Autonomous self-pollination is defined as the transfer of pollen from the anther to the stigma of the same flower (Bateman 2020, Johnson & Edwards 2000, Willmer 2011). Several studies have shown that orchids can be pollinated with their own pollen and produce fruits and viable seeds (Ackerman *et al.* 2023). However, the fact that a plant could be self-pollinated does not necessarily make it obligatorily autogamous, as the majority of orchids commonly show different degrees of facultative autogamy (Talalaj *et al.* 2017). In contrast, allogamous self-compatible species that can be self-pollinated manually are called "frequently cross-pollinated species" (Sasikumar 2010). This knowledge is widely applied in *V. planifolia*, an orchid species self-pollinated by hand (Hernández-Hernández 2018). Autogamy guarantees the production of fruits, if that is what is desired, even knowing that the seeds contained may not be viable (Sao Leao *et al.* 2019, Yeh *et al.* 2021). Another disadvantage observed in self-pollinated orchids is the production of albino protocorms, something not commonly observed in orchids pollinated through outcrossing (de Paiva-Neto *et al.* 2022). Endogamy depression can be expressed as auto-pollination or in crosses between closely related individuals, as reported by Emeterio-Lara *et al.* (2018).

Although orchid flowers are widely adapted to outcrossing, species capable of autogamy are found in several Orchidaceae subgroups (Johnson & Edwards 2000). Spontaneous self-pollination is relatively common in Orchidaceae compared to other plant families. In fact, 31 % of orchid species set fruits through autonomous self-pollination (Evans & Jacquemyn 2020). The occurrence of self-pollination is easy to study in orchids due to its floral architecture (Talalaj *et al.* 2017). In addition, facultative self-pollination is sometimes observed in biotic-pollinated plants when the environment is subject to changes such as anthropogenic disturbance (Talalaj & Skiercynski 2015, Talalaj *et al.* 2017). This transition has also been recorded in some plants that have migrated to areas outside their previous range (Sramkó *et al.* 2019, Evans & Jacquemyn 2020). However, autogamy is more common among orchids with a weedy habit or those found in habitats with marginal pollinator activity (Suetsugu 2015).

Orchids developed barriers to avoid the production of seeds with self-pollen, and these can occur in both before and after pollen transfer. Autogamy in orchids is often prevented or limited by various mechanisms to promote outcrossing, which enhances genetic diversity and consequently more adaptability. Some common barriers to autogamy in orchids are 1) Morphological barriers: in many orchid species, the anther and stigma are spatially separated by the rostellum, preventing self-pollination (Bory et al. 2008, Sugiura 2013, Zhang et al. 2021). 2) Temporal barriers: some orchids exhibit dichogamy, where the male and female reproductive organs mature at different times; this temporal separation prevents self-pollination because the receptive stigma is not vet available when pollen is released or vice versa (Hurskainen et al. 2017). 3) Genetic barriers: some orchid species possess mechanisms of self-incompatibility, where the pollen from a flower is unable to fertilize the ovules of the same flower or other flowers on the same plant; this genetic barrier prevents self-fertilization and promotes outcrossing (Zhang et al. 2021). Genetic barriers such as self-incompatibility, operate after pollinia transfer by ensuring that the pollen from one flower cannot successfully fertilize the ovules of the same flower or other flowers on the same plant (Hurskainen et al. 2017, Zhang et al. 2021). 4) Other mechanical barriers: orchids typically have specialized pollination structures associated with the pollinia or pollinaria, which are often adapted for specific pollinators; these structures may not easily come into contact with the stigma of the same flower, reducing the likelihood of self-pollination (Gravendeel et al. 2004). Overall, these barriers to autogamy in orchids contribute to the promotion of outcrossing, which facilitates genetic recombination and maintains genetic diversity within populations (Scopece et al. 2014, Suetsugu 2015, Zhang et al. 2021).

Often, autogamy in orchids is facultative, and self-pollination takes place in the final phase of anthesis, mainly caused by a low frequency of pollinator visits (Suetsugu 2015, Talalaj *et al.* 2017). However, self-pollination becomes mandatory when the pollinator is ineffective or is no longer in the area (Jin *et al.* 2014, Pedersen *et al.* 2018). For example, *V. palmarum* (Salzm. Ex Lindl.) Lindl. opens its flowers and remains available to pollinators, however, it also exhibits a mechanism of facultative self-pollination; if biotic pollination does not occur, flowers self-pollinate (Pansarin & Ferreira 2021).

The transition between allogamy-autogamy pollination systems is particularly well documented in the terrestrial genus *Epipactis* (Talalaj *et al.* 2017, Sramkó *et al.* 2019, Evans & Jacquemyn 2020). *Epipactis* flowers show a morphology adapted to self-pollination and ensure seed production such as a reduced gynostemium or the occurrence of cleistogamy ensure seed production. Autogamy has been recorded in *E. atrorubens* (Hoffm. Ex Bernh.) Besser, *E. dunensis* Godfery, *E. helleborine* (L.) Crantz, *E. leptochila* Godfery, *E. microphylla* (Ehrh.) Sw., *E. muelleri* Godfery, *E. palustris* (L.) Crantz, and *E. youngiana* A.J.Richards ex A.F.Porter (Bonatti *et al.* 2006, Talalaj & Brzosko 2008, Talalaj *et al.* 2017, Evans & Jacquemyn 2020). Nonetheless, this adaptation has reduced the genetic diversity of the populations (Squirrell *et al.* 2002, Evans & Jacquemyn 2020). Even though the transition from allogamy to autogamy has been considered as an evolutionary dead end, due to the consecutive acquisition of individuals with lower genetic diversity, and therefore, with less probability of developing mechanisms that allow them to adapt to environmental changes, which might lead to narrow the ecological niches and eventually the loss of the species, this is common in orchids, since they have strategies to persist (*i.e.* clonality) (Phillips *et al.* 2020, Stipková *et al.* 2020, Evans & Jacquemyn 2023).

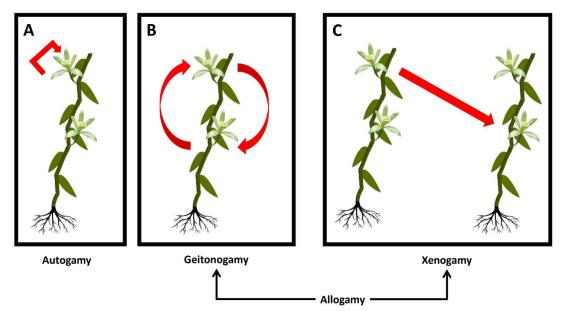


Figure 2. Comparison between autogamy (A) and allogamy (geitonogamy (B) and xenogamy (C)). Red arrows indicate the direction from pollen donator flower to pollen receipt flower. The diagram represents the species *Vanilla planifolia*, flowers are not in real scale.

Allogamy. It is subdivided in two types: geitonogamy (the pollen is taken to the stigma of another flower, from the same individual), and xenogamy (the pollen is taken to the stigma of another flower, from a different individual) (Bateman 2020, de Oliveira *et al.* 2022, de Paiva-Neto *et al.* 2022). A graphical comparison between autogamy, geitonogamy and xenogamy is shown in Figure 2.

Due to the structure of their flowers, allogamy is the most common breeding system among orchids (Lanzino *et al.* 2023). Even Darwin focused on explaining mechanisms that could promote cross-pollination, as he thought that such floral adaptations of the orchid were selected as consequence of pollinator pressures (Scopece *et al.* 2014). Orchid flowers commonly exhibit mechanical barriers (*i.e.*, rostellum) that tends to reduce the formation of fruits with self-pollen (Zhang *et al.* 2021). This tendency to avoid self-pollination in orchids is due to the fact that self-incompatibility is a post-pollination barrier that avoids self-fertilization, however, self-incompatibility does not avoid pollen transfer (pollination) (Valdivia *et al.* 2010, Zhang *et al.* 2021). In partially self-incompatible orchids commonly occur a reduction in the number of seeds per fruit, reduction of the weight and size of the fruits, and a lower number of seeds with embryo (Tremblay *et al.* 2005, de Paiva-Neto *et al.* 2022). Partial self-incompatibility has been recorded in more than 750 species of Orchidaceae, mainly in *Chondrorhyncha, Coelogyne, Dendrobium, Lycaste, Notylia* and *Oncidium* (Johnson & Edwards 2000, Tremblay *et al.* 2005).

The first case of allogamy discussed here is geitonogamy. Almost all flowers from an inflorescence usually have the same probability of being visited by a pollinator, thus, geitonogamy results in the same effort as observed in xenogamy, because pollen is transported between two flowers (Lanzino *et al.* 2023). However, geitonogamy has been considered similar to autogamy, as the pollen transfer occur between flowers of the same individual (Kropf & Renner 2008). For this reason, some authors have considered that, in order to increase genetic diversity, geitonogamy is undesirable, along with autogamy (Sletvold *et al.* 2012, Gigant *et al.* 2016). Besides, it must be considered that there are many orchids that produce solitary flowers or racemes whose flowers open successively (one after another), which may reduce the probability of geitonogamy (Srimuang *et al.* 2010). Nevertheless, in some orchid species such as *Chloraea crispa* Lindl., *Eulophia alta* (L.) Fawc. & Rendle, or *Phaius tankervilleae* (King & Pantl.) Karthik., pollination by geitonogamy results in higher fruit set and seed germination than by autonomous self-pollination (Humaña *et al.* 2008, Johnson *et al.* 2009, Buragohain *et al.* 2016), in comparison to autogamy or even xenogamy, considered as the best way of pollinating (discussed below).

In contrast, pollination by xenogamy is the most widespread type of allogamy in orchids, which is facilitated by some floral structures such as the rostellum (Bory *et al.* 2008, Sugiura 2013, Freudenstein & Chase 2015, Ospina-Calderón *et al.* 2015). In truly allogamous orchids this structure is well-developed, while in autogamous species it is usually reduced (Suetsugu 2015, Endress 2016). Although today we know that xenogamy is not the only form of pollination, previous works, such as Darwin's statement, mentioned that it must be the main way (Scopece *et al.* 2014).

There is a tendency observed in allogamous plants (not only in orchids) to be pollinated by xenogamy instead of geitonogamy (Pang & Saunders 2015, Kundu & Karmakar 2022). It is hypothesized that the lack of nectar evolved to reduce geitonogamy, because pollinators (mainly bees), tend to avoid non-rewarding flowers (Johnson *et al.* 2004, Kropf & Renner 2008). A consequence of xenogamy is the genetic variation, allowing a better adaptation than to obligatory autogamous species (Willmer 2011). In addition, it provides some defense against natural mutations in the genetic material, since if one of the nuclear genomes is damaged, the effects of non-functional alleles may be masked by the correct functioning of the equivalent alleles on the chromosome inherited from the other parent (Willmer 2011, Talalaj & Skiercynski 2015). Autogamy reduces genetic diversity and favors the expression of harmful genes (Willmer 2011, Emeterio-Lara *et al.* 2018).

Pollination by xenogamy can also influence both the fruit set and the seed production (Tremblay *et al.* 2005). Even in self-compatible plants, this mode of pollination commonly increases fruit set (Borba *et al.* 1999, Caballero-Villalobos *et al.* 2017, Emeterio-Lara *et al.* 2018, Sao Leao *et al.* 2019), and improves seed viability (Johnson 2000, Vale *et al.* 2010, Caballero-Villalobos *et al.* 2017, Capó *et al.* 2022). Several studies have shown that allogamy generally tends to be more advantageous compared to autogamy in terms of higher fruit set, as observed in *Epidendrum denticulatum* Barb. Rodr. (Sao Leao *et al.* 2019), or *Vanilla palmarum* (Salzm. Ex Lindl.) Lindl. (Pansarin & Ferreira 2021), species that exhibited a fruit set between 43-76 % through autogamy, and 82-95 % through allogamy. This advantage stems from the genetic benefits associated with outcrossing, such as increased genetic diversity and reduced risk of inbreeding depression (Valdivia *et al.* 2010, Zhang *et al.* 2021). Cross-pollination increases the chances of successful fertilization by introducing genetic diversity and potentially overcoming self-incompatibility mechanisms (Zhang *et al.* 2019). Also, xenogamy generally results in seeds with higher viability compared to autogamy, as recorded in *Cattleya coccinea* Lindl. (Caballero-Villalobos *et al.* 2017), or *Cyrtopodium punctatum* (L.) Lindl. (Dutra *et al.* 2009), since these species exhibited a germination percentage ranging from 9 to 65 % through autogamy, and 42 to 97 % through xenogamy.

Many orchid species set fruits exclusively by cross-pollination (Fantinato *et al.* 2017, Mosquera-Mosquera *et al.* 2019, Zhang *et al.* 2021). Epidendroideae orchids exhibit a greater number of self-sterile species, and few Orchidoideae orchids can be self-fertilized (Fantinato *et al.* 2017, Sao Leao *et al.* 2019, Zhang *et al.* 2021). In contrast, orchids from Vanilloideae and Cypripedioideae species can set fruits through self-pollination, although their floral morphology promote cross-pollination (Suetsugu & Fukushima 2014, de Oliveira *et al.* 2022). Finally, Apostasioideae orchids might be pollinated by xenogamy (Kocyan-& Endress 2001, Yin *et al.* 2016). A comparison of pollination systems among different orchid subfamilies is shown in Table 1.

If pollination by allogamy (mainly by xenogamy) exhibits so many advantages over autogamy, why are there species that remain autogamous? In true autogamy can be an important sexual process of reproduction in flowering plants. Although it is not as effective as xenogamy, pollination by autogamy might exhibit a certain degree of genetic recombination (Willmer 2011). Additionally, autogamy does not require a substantial energy investment in the reward production (Eckert & Herlihy 2004). The participation of pollinators is no longer necessary, which makes pollination by autogamy advantageous in the case of pollinator decline (Pedersen & Ehlers 2000). Many orchids, especially in the tropics, show populations with few individuals, and successful cross-pollination events can be rare (Bernhardt & Edens-Meier 2010, Cabrera-Reyes *et al.* 2021). A species under such circumstances would develop strategies to ensure its offspring production, where autogamy would serve as a mechanism for this, albeit at the expense of potential adverse effects as mentioned above. However, cross pollination prevails throughout most members of this family and autogamy only happens as a last resort, which could mean orchids prioritize the advantages of

Subfamily	Main conclusions	Cites
Apostasioideae	There are not formal studies about pollination ecology within this sub- family. It is hypothesized that pol- lination might occur by xenogamy, and pollen grains (pollen is not agglutinated) might be offered as a reward to their pollinator, possibly meliponini bees.	Kocyan & Endress 2001, Yin <i>et al.</i> 2016
Vanilloideae	Although historically hand-polli- nated through autogamy, the genus <i>Vanilla</i> displays physical barriers (rostellum) on the flower, suggest- ing a predisposition to pollination by outcrossing rather than autoga- my. The natural pollinators (when available) are euglossini bees. There is a single anther, with agglutinated pollen which is released as monads or tetrads.	Sasikumar 2010, Pansarin & Ferreira 2021, de Oliveira <i>et</i> <i>al.</i> 2022
Cypripedioideae	Most of the species are self-compat- ible, but their flowers are designed as one-way traps, and because of their morphology, they promote cross pollination. They require in- sects (mainly wild bees) to transfer pollen from the anther to the stigma. There are two fertile anthers, whose pollen is agglutinated and paste-like.	Pemberton 2013, Suetsugu & Fuku- shima 2014
Orchidoideae	It presents authentic pollinia, but divisible. There are some self- compatible species, they do not exhibit a rostellum, and if they do, it is very narrow or degrades during floral opening. However, xenogamy is considered as the predominant way of pollinating. Pollinators are mainly bees and lepidoptera.	Pansarin & Ferreira 2015, Fantinato <i>et al.</i> 2017
Epidendroideae	It presents authentic pollinia, and it is indivisible. Xenogamy is consid- ered as the predominant way of pol- linating since self-compatibility is exhibited in most of its species. Epi- dendroideae orchids have often been considered to be nectar-rewarding or nectarless, however, some few species reward with nectar to their pollinators. Pollinators are flies and lepidoptera, or even hummingbirds.	Pansarin & Pansarin 2016, Mosquera- Mosquera <i>et al.</i> 2019, Sao Leao <i>et al.</i> 2019, Zhang <i>et al.</i> 2021

Table 1. Comparison among the different subfamilies of Orchidaceae regarding main breeding s	system exhibited.
--	-------------------

cross-pollination over the potential security of fruit and seed production through autogamy, which does not ensure the production of viable seeds (Sao Leao *et al.* 2019, Yeh *et al.* 2021, de Paiva-Neto *et al.* 2022).

Metaxenia and xenia. All pollination strategies previously mentioned involve only one species (intra-specific pollination). Conversely, in metaxenia and xenia two species are involved (inter-specific pollination). Metaxenia is understood as the effect of foreign pollen (different species, but from the same genus) on fruit formation, including size increase and changes in traits as texture, shape, scent, flavor, and chemical composition (MacInnis & Forrest 2020). In addition, biochemical changes are also presented, as an increase in the concentration of different metabolites, highlighting carbohydrates and phenolic compounds (Suaib *et al.* 2020, Shahsavar & Shahhoseini 2022).

Although there is no explanation regarding the molecular mechanisms related to metaxenia, current hypotheses point out to an increase in the concentration of growth regulators such as auxin, cytokinin, and gibberellin from the new pollen source, as fruits of larger size in other cultivars tend to exhibit higher levels of these regulators (Cheng *et al.* 2020, Deng *et al.* 2022). Another hypothesis suggests an enzymatic change in the fruits, particularly those enzymes related to processes such as fruit expansion and the production of sugars and phenolic compounds (Deng *et al.* 2022, Shahsavar & Shahhosseini 2022). Besides, chemical signaling related to volatile compounds could be another plausible explanation, since some chemical signals produced by the male part of the flower (released by the pollen) may interact with female reproductive structures and influence fruit development (Piotto *et al.* 2013, Deng *et al.* 2022). These hypotheses have been proposed based on observations in other crops of importance (*i.e.*, date, pear, plum and tomato); however, these postulates have not been verified in orchids or in wild plants.

A related concept is xenia, sharing the same definition mentioned for metaxenia, but applied to seeds (Sabir 2014). It has been reported that xenia seeds usually exhibit greater viability, manifested as greater germination and development of seedlings, as well as an increase in size and weight (Sattler *et al.* 2016, van Esse *et al.* 2020). Xenia is considered the previous step for the formation of hybrids, since compatibility between the species must be observed, in addition to observing the immediate effects of the pollen (Sari *et al.* 2023). It has been considered that these changes in the seed could be attributable to greater control of seed development by paternal genes, and that maternal genes may lose part of the control, by allowing the formation of large seeds (de Jong & Scott 2007). However, this hypothesis was proposed for other angiosperms but the mechanisms of xenia induction in orchids are still unknown.

The study of metaxenia and xenia emerged in the last century, with the study carried out by Swingle (1928) focused on the date palm. Metaxenia as a concept is relatively recent, considering that the study of pollination ecology formally began in 1793 (Faegri & Pijl 1979). Studies about the effect of xenia and metaxenia induction have focused mainly on crops of economic importance, such as apple (Bodor *et al.* 2008, Militaru *et al.* 2015), pear (Cheng *et al.* 2020), strawberry (MacInnis & Forrest 2020), date (Swingle 1928, Shahsavar & Shahhoseini 2022), tomato (Piotto *et al.* 2013), cucumber (Olfati *et al.* 2010), grape (Sabir 2014), hazelnut (Balik & Beyhan 2020), and corn (Suaib *et al.* 2020), among others. In general, a positive effect could be observed regarding fruits and seeds production. Therefore, it is recommended to carry out this breeding system to obtain improvements in crops.

No formal studies about xenia and metaxenia have been carried out in Orchidaceae, except for two subfamilies: two species from Vanilloideae (*V. planifolia* and *V. pompona* Schiede) (Menchaca-García *et al.* 2011, Barreda-Castillo *et al.* 2023a), and three species from Epidendroideae (*Bulbophyllum weddellii* (Lindl.) Rchb.f, *B. involutum* Borba, Semir & F. Barros, and *B. ipanamense* Hoehne) (Borba *et al.* 1999). Regarding the *Vanilla* species, interspecific pollinations were carried out since both vanillas coincide phenologically (Barreda-Castillo *et al.* 2023a). A beneficial effect due to the induction of metaxenia could be observed only in *V. planifolia* when was pollinated with *V. pompona*, manifested as an increase in size and weight in the fruits in comparison to autogamy in each species, while in *V. pompona* it was the opposite effect, presenting smaller fruits. So, although it is beneficial in most crops, the induction of metaxenia does not necessarily provide better results compared to traditional pollination. Moreover, Menchaca-García *et al.* (2011) reported xenia effect on both *V. planifolia* and *V. pompona* species, obtaining germination percentages close to 80 %, which means an increase, considering that *V. planifolia* presents germination percentages near to 5 % (Yeh *et al.* 2021), and *V. pompona* germination has not been reported (Menchaca-García *et al.* 2011). This study was originally called "hybrid production", however, xenia and hybrid production are related concepts, as we discussed in the next section.

Metaxenia and xenia induction are related to interspecific hybrids production, something useful to the most commercially important orchid, and one of the highest yielding crops in the tropics, *Vanilla*. It is recommended, first of all, to continue searching for pollen donor species for *V. planifolia*, aiming to induce changes in the chemical composition of its fruits, as well as an increase in their size (Chambers 2019). In addition, hybrid organisms would be obtained indirectly, combining qualities of the parental species (Sari *et al.* 2023). Furthermore, by selecting pollen donor species, these could accompany the cultivation of *V. planifolia*, which in turn could contribute to breeding programs for vanilla species (Watteyn *et al.* 2023) or even species with agro-economic value.

Regarding the *Bulbophyllum* species, crosses between *B. involutum* and *B. ipanamense* (species with greater genetic proximity) exhibited fruit formation rates and seed viability similar to those obtained in intraspecific crosses, whereas crosses between *B. weddellii* with both *B. involutum* and *B. ipanamense* exhibited a higher rate of fruit abortion (Borba *et al.* 1999). Therefore, better results are obtained in metaxenia and xenia induction when the species reflect genetic proximity. This study was reported as "crossing potential", although they actually reported metaxenia and xenia effect. Research on the effect of xenia and metaxenia in orchids is necessary, given the lack of information that currently exists.

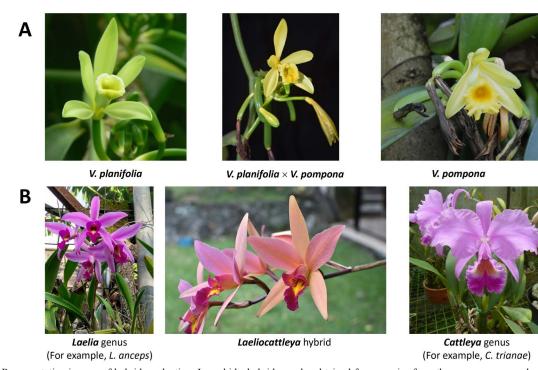


Figure 3. Representative images of hybrid production. In orchids, hybrids can be obtained from species from the same genus, as observed in *Vanilla* planifolia × V. pompona (A), or different genera, as observed in *Laeliocattleya* (B). L. anceps and C. trianae are illustrative only.

Hybrids. Although the effect of xenia induction has not been properly studied in orchids, the capacity of hybridization is well known in this plant family. A hybrid is defined as the organism result of cross fertilization between different species from the same genus (Figure 3A) or between species belonging to distinct genera (Figure 3B) (Kishor & Sharma 2009, Kempe & Gils 2011, López-Caamal & Tovar-Sánchez 2014, Chambers 2019). Despite metaxenia and xenia induction along with hybrids production involve cross pollination, metaxenia effect is just observed in fruit

formation (Balik & Beyhan 2020), xenia effect is related to seed production (Sabir 2014), whereas hybrid production relates to the new generation of organisms produced (Preston & Pearman 2015, Goulet *et al.* 2017).

Xenia is directly related to hybrid production, as xenia induction is a process in plant reproduction that involves manipulating pollination to produce specific characteristics in the offspring (Olfati *et al.* 2010). In the context of hybrid production, xenia induction is used to ensure the fertilization of an ovule by a specific pollen grain, resulting in a hybrid with desirable traits from both parents (Deng *et al.* 2022). In artificial hybridization, this process is used to combine the desirable traits of two different parental entities on the hybrid (Goulet *et al.* 2017). For example, if a hybrid with greater resistance to certain diseases and improved performance is desired, breeders can use xenia induction to ensure that the ovules of one plant are fertilized by the pollen of another plant with the desired characteristics.

On the other hand, interspecific and intergeneric hybrids are related to the theory of metaxenia or xenia in the context of plant breeding and reproduction. Interspecific hybrids are produced by crossing between two different species within the same genus; in the context of metaxenia or xenia, when these hybrids are formed, characteristics from both parent species can be influenced by xenia, leading to traits in the offspring that are influenced not only by the genetic makeup of the parents but also by the environmental effects on the maternal tissues surrounding the embryo sac during pollination and fertilization (de Jong & Scott 2007, Malaviya *et al.* 2019, Deng *et al.* 2022). In contrast, intergeneric hybrids are produced by crossing two different genera; similarly, in the context of metaxenia or xenia, when intergeneric hybrids are formed, the environmental effects on the maternal tissues during pollination and fertilization and fertilization (de Jong & Scott 2007, Malaviya *et al.* 2019, Deng *et al.* 2022). In contrast, intergeneric hybrids are produced by crossing two different genera; similarly, in the context of metaxenia or xenia, when intergeneric hybrids are formed, the environmental effects on the maternal tissues during pollination and fertilization can influence the traits of the resulting offspring, along with the genetic contributions from both parent genera (Havkin-Frenkel & Belanger 2018, Li *et al.* 2021a, Vilcherrez-Atoche *et al.* 2022).

In the past, it was considered that the production of plant hybrids was useless, since it would not be possible to obtain viable organisms, and if any were obtained, they would be weak and inferior to their parental species (Stebbins 1958). Nowadays, it is known that the hybridization is a viable process observed in several plant groups (Paun *et al.* 2011), as it occurs even in natural conditions (Fay *et al.* 2007, Johnson 2018, Arida *et al.* 2021, Cantuária *et al.* 2021).

Natural hybridization in orchids have significant contributions in evolutionary, ecological, and taxonomic processes. Regarding evolutionary processes, natural hybridization is a common phenomenon and has long been suspected to be a potent evolutionary force (Li *et al.* 2021a, Fiorini *et al.* 2023). It has been suggested that a significant number of flowering plants may be of hybrid origin (Cozzolino *et al.* 2006). Orchid hybrids contribute to the ongoing evolutionary processes by introducing new genetic combinations and variations (Cozzolino *et al.* 2006, Johnson 2018, Evans *et al.* 2023). Also, orchid hybrids can play a role in speciation processes by serving as intermediates between parental species (Johnson 2018). Hybridization events can lead to the formation of new species through hybrid speciation, where hybrids become reproductively isolated from parental species and establish distinct evolutionary lineages (Fay *et al.* 2007, Pavarese *et al.* 2013, Marques *et al.* 2014). Hybridization is not merely a kind of "evolutionary noise" with little evolutionary significance but may instead sometimes play a positive role in evolution, either through hybrid speciation, or through the origin and transfer of novel adaptations (Cozzolino *et al.* 2006).

Natural hybridization is really common in Orchidaceae (Johnson 2018, Evans *et al.* 2023), as it has been observed in *B.* × *cipoense* (*B. weddellii* × *B. involutum*) (Borba & Semir 1998), *Catasetum* × *sheyllae* (*C. boyi* Mansf. × *C. garnettianum* Rolfe) (Cantuária *et al.* 2021), *Epidendrum* × *purpureum* (*E. denticulatum* × *E. orchidiflorum* Salzm. ex Lindl.) (Arida *et al.* 2021), *Laelia* × *meavei* (*L. dawsonii* (J.Anderson) De B. Crawshay × *L. rubescens* Lindl.) (Cetzal-Ix *et al.* 2020), *L.* × *oaxacana* (*L. halbingeriana* Salazar & Soto Arenas × *L. anceps* Lindl.) (Salazar *et al.* 2014), *Prostechea* × *chixoyensis* (*P. cochleata* (L.) W.E.Higgins × *P. radiata* (Lindl.) W.E.Higgins) (Mó *et al.* 2014), *Orchis* × *dietrichiana* (*O. tridentata* Scop. × *O. ustulata* L.) (Cozzolino *et al.* 1998), or *Vanilla* × *tahitensis* (*V. planifolia* × *V. odorata* C.Presl), which was believed to be a species (Lubinsky *et al.* 2008), among others.

Regarding ecological processes, natural orchid hybrids can potentially reproduce with their parent species, leading to introgressive hybridization (Marques *et al.* 2014). This process involves the transfer of genetic material between hybrids and parent species, influencing the genetic diversity and adaptation potential of populations (Marques *et al.* 2014).

2014). Introgressive hybridization can facilitate the exchange of adaptive traits, contributing to the resilience and evolutionary flexibility of orchid populations in changing environments (Pinheiro *et al.* 2010). However, natural hybridization is typically considered deleterious for the conservation of biodiversity (Vereecken *et al.* 2010, Stull *et al.* 2023). Interspecific gene flow is often seen as a hazard in plant conservation genetics, especially when rare species come in contact and hybridize with more common and widespread related taxa as a consequence of habitat disturbance (Ferdy & Austerlitz 2002). Hybridization may lead to the loss of rare taxa as a consequence of outbreeding depression and genetic assimilation (Chung *et al.* 2005). Consequently, specific conservation strategies should be designed to protect individuals and hybrid populations, in order to maintain both the natural sources and the new organisms for future evolution (Fay 2018, Evans *et al.* 2023).

Regarding taxonomic contributions, natural orchid hybrids contribute to the overall diversity of the orchid family by generating new combinations of traits and morphologies (Goulet *et al.* 2017, Li *et al.* 2021a). These hybrids often exhibit unique characteristics that may not be present in either parent species, leading to the recognition of additional taxa (Bertrand *et al.* 2021). However, orchid hybrids can also present challenges for taxonomic classification due to their intermediate characteristics and complex genetic backgrounds (Cozzolino *et al.* 2006, Radak *et al.* 2019). Therefore, taxonomists need to carefully evaluate morphological, genetic, and ecological data to accurately classify orchid hybrids and understand their evolutionary relationships with parent species.

In addition to natural hybridization, there are several hand-made hybrids between species of different genera, for example, *Aranda (Arachnis hookeriana* (Rchb. f.) Rchb. f. × *Vanda lamellata* Lindl.), *Brassocattleya (Brassovola* × *Cattleya), Laeliocattleya (Laelia* × *Cattleya), Odontocidium (Odontoglossum* × *Oncidium)*, and several interspecific hybrids within *Cattleya, Cymbidium, Dendrobium* or *Phalaenopsis* (Asociación Mexicana de Orquideología 2005). In orchids, the hybrid production is not limited to primary hybrids, since there are secondary hybrids (crossing of a hybrid with a species or crossing three species) reported (*i.e., Vanilla* × *manitra ampotony (V. planifolia* × *V.* × *tahitensis*)) (Grisoni & Nany 2021). In addition, hybrids can be cross-pollinated with other hybrids, and viable seedlings can still be obtained, combining more traits in each generation (Devadas *et al.* 2016).

Among the 25 largest plant families, Orchidaceae is the one with the largest number of known hybrids (Fiorini *et al.* 2023). The lack of endosperm and the phylogenetic closeness that orchids share with each other have been considered as the main reasons why so many hybrids are obtained in this family (Johnson 2018, Li *et al.* 2021a). Although phenological coincidence is helpful in the formation of hybrids because pollinators might cross-pollinate between species (Turchetto *et al.* 2022), as well geographical coincidence, also called "hybrid zones" (Marques *et al.* 2014, Johnson 2018, Evans *et al.* 2023), and morphological similarity of flowers (Calevo *et al.* 2021). However, sometimes pollen is preserved in cryogenic conditions to be used later, achieving both formation of fruits as well as viable seeds (Divakaran *et al.* 2016), but this happens under cultivation conditions or in horticulture.

The first list of hybrid orchids reported around 10,000 crosses (Adams & Anderson 1958). The first natural orchid hybrid ever recorded was *Phalaenopsis intermedia* (*P. aphrodite* Rchb. f. × *P. equestris* var. *rosea* Valmayor & D. Tiu) in 1853 (Li *et al.* 2021a), whereas the first hand-made orchid hybrid was "Calanthe" (*Calanthe masuca* (D. Don) Lindl. × *C. furcata* Bateman ex Lindl.), reported in 1856 (Li *et al.* 2021a). Nowadays, more than 100,000 orchid hybrids (natural and hand-made) are reported worldwide (The Royal Horticultural Society 2023), list that keeps growing every day.

In most of the cases, hybridization process results in the production of organisms with better responses to biotic and abiotic stress, in comparison to their parental species (Divakaran *et al.* 2006, Kumar & Singh 2016, Goulet *et al.* 2017, Li *et al.* 2021a). Some benefits of the production of hybrid organisms are reflected in qualities appreciated by the market, such as better blooms, larger flowers, long flowering period, or the combination of pigments (Tatsuzawa *et al.* 2004, Edens-Meier *et al.* 2013, Pramanik *et al.* 2022). Perhaps this is the main reason why there are so many artificial orchid hybrids worldwide.

Improvement of shape, size and aroma of the fruit might not be important for most orchids; however, it is very important in the case of vanilla. For this reason, research on hybrids in orchids is of greatest relevance in vanilla,

due to its economic importance. For example, *V. planifolia* × *V. pompona* hybrids have exhibited a better response to water stress (Barreda-Castillo *et al.* 2023b), and to the exposition to *Fusarium oxysporum* f. sp. *vanillae*, its main pathogen (Barreda-Castillo *et al.* 2022). *V.* × *tahitensis* (*V. planifolia* × *V. odorata*) exhibits a vanillin concentration similar to *V. planifolia* (Brunschwig *et al.* 2016). *V.* × *manitra ampotony* (*V. planifolia* × *V.* × *tahitensis*) shows a concentration of vanillin (aromatic chemical marker of vanilla) up to 20 times higher than the observed in *V. planifolia*, in addition to an increase in certain phenolic compounds (Grisoni & Nany 2021). *V.* × *tsy taitra* ((*V. planifolia* × *V. pompona*) × *V. planifolia*) exhibits a better aromatic quality than *V. planifolia*, along with the resistance to *F. oxysporum* f. sp. *vanillae* (Varela-Quirós 2010, Havkin-Frenkel & Belanger 2018). More vanilla hybrids have been produced between aromatic species, such as *V. planifolia* × *V. phaeantha* Rchb. f., *V. phaeantha* × *V. pompona*, or *V. pompona* × *V. odorata* (Hu *et al.* 2019, Chambers *et al.* 2021), with the aim of obtaining higher quality fruits.

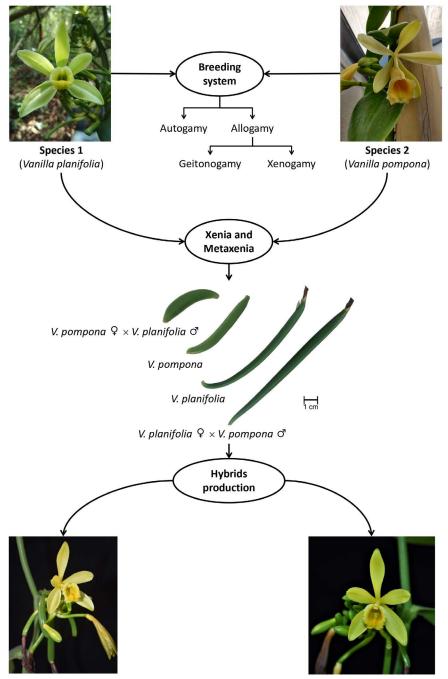
Even though historically so many hybrids have been produced within Orchidaceae, not all of them have been viable, or have shown optimal characters (Chambers 2019, Grisoni & Nany 2021). Hybrid production programs as a genetic improvement strategy have historically produced hundreds of lines, however, only certain lines of interest have been selected (Grisoni & Nany 2021). It is recommended to select the orchid hybrids since seed germination (Menchaca-García 2018), or during the development of seedlings (Divakaran *et al.* 2016), in order to conserve the lines with the desired characters, and once selected, multiply them by *in vitro* culture (Divakaran *et al.* 2016, Menchaca-García 2018). Although genetic improvement of orchids through hybridization is a long-term process, it is still the best method to obtain plants with improved qualities without having to resort to the use of transgenics, since hybrid organisms are not necessarily result of genetic modification (Chandler & Dunwell 2008, Goulet *et al.* 2017, Chambers 2019).

Finally, it is recommended that in future hybridization programs the species with the most desirable characters must be used as ovule donor instead of the pollen donor, due to greater expression of maternal traits in the new organisms (Havkin-Frenkel & Belanger 2018, Barreda-Castillo *et al.* 2023b). There are two main hypotheses about the expression of maternal characters in a greater degree in hybrids: 1) It might be due to the inheritance of plastids and mitochondria genomes, since both organelles are usually inherited from the maternal parent (Daniell *et al.* 2021, Park *et al.* 2021) or 2) it might be due to epigenetic regulation, since this mechanism is more sensitive in plants (Baulcombe & Dean 2014, Kumar & Singh 2016). However, there is not a real consensus about this topic. All types of pollination mentioned so far (as well as the induction of metaxenia and xenia, and production of hybrids) are summarized in Figure 4.

Members of Orchidaceae offer several types of rewards to pollinators or are pollinated by any kind of deception, but fruit set by cross-pollination is strongly favored in this family. However, obligatory and facultative autonomous self-pollination has evolved independently several times in this huge plant family. Allogamy is favored by pre-zygotic, *i.e.*, floral mechanisms, and post-zygotic (genetic) barriers. Moreover, pollination by xenogamy is more frequent than by geitonogamy, because xenogamy offers benefits such as greater genetic diversity.

Although Orchidaceae is well known for its capacity for interspecific pollination and viable production of fruits and seeds, there are scarcity of studies about biochemical and morphophysiological changes in fruits (metaxenia) or seeds (xenia) in these species. Paradoxically, hybrid production (result of interspecific pollination) is well documented in this family, since Orchidaceae shows the greatest production of these organisms. It is recommended to keep producing hybrids for ornamental and economical purposes, as increasing crop productivity, as in the case of *Vanilla*. Besides, these organisms usually exhibit better traits than their parental species, expressed as greater tolerance to adverse conditions such as biotic or abiotic stress.

In summary, although the majority of orchids can be self-pollinated, it is advisable to pollinate them through xenogamy when done by hand, since in this way it is possible to promote intraspecific genetic diversity and promote seed germination, something necessary in species in risk of extinction. Furthermore, induction of xenia (and hybrid production) could promote genetic improvement, and production or organisms with new and desirable traits.



V. planifolia $\mathcal{Q} \times V$. pompona \mathcal{O}

V. pompona $\mathcal{Q} \times V$. planifolia \mathcal{O}

Figure 4. Breeding systems, induction of xenia and metaxenia, and production of hybrids in orchids. Breeding systems autogamy or allogamy (geitonogamy and xenogamy) involves only one species, whereas xenia and metaxenia require two. Although xenia and metaxenia effect only is associated with seeds and fruits production, respectively, hybrids production is related to this process, as the new organisms are also result of cross pollination.

Acknowledgements

We thank to M. Lozano Rodriguez, E. Flachland and J. Santos for sharing several orchid photos used in this article. Authors are grateful to the reviewers and the editor of section for their comments.

Literature cited

- Ackerman JD, Phillips RD, Tremblay RL, Karremans A, Reiter N, Peter CI, Bogarín D, Pérez-Escobar OA, Liu H. 2023. Beyond the various contrivances by which orchids are pollinated: global patterns in orchid pollination biology. *Botanical Journal of the Linnean Society* 202: 295-324. DOI: <u>https://doi.org/10.1093/botlinnean/boac082</u>
- Adams H, Anderson E. 1958. A conspectus hybridization in the Orchidaceae. *Evolution* **12**: 512-518. DOI: <u>https://doi.org/10.2307/2405962</u>
- Anghelescu NE, Balogh M, Balogh L, Kigyossy N, Georgescu MI, Petra SA, Toma F, Peticila AG. 2023. *Epipactis bucegensis*-A separate autogamous species within the *E. helleborine* Alliance. *Plants* 12: 1761. DOI: <u>https://doi.org/10.3390/plants12091761</u>
- Antonelli A, Fry C, Smith RJ, Eden J, Govaerts RHA, Kersey P, Nic Lughadha E, Onstein RE, Simmonds MSJ, Zizka A, Ackerman JD, Adams VM, Ainsworth AM, Albouy C, Allen AP, Allen SP, Allio R, Auld TD, Bachman SP, Baker WJ, Barrett RL, Beaulieu JM, Bellot S, Black N, Boehnisch G, Bogarín D, Boyko JD, Brown MJM, Budden A, Bureš P, Butt N, Cabral A, Cai L, Cano JA, Chang Y, Charitonidou M, Chau JH, Cheek M, Chomicki G, Coiro M, Colli-Silva M, Condamine FL, Crayn DM, Cribb P, Cuervo-Robayo AP, Dahlberg A, Deklerck V, Denelle P, Dhanjal-Adams KL, Druzhinina I, Eiserhardt WL, Elliott TL, Enquist BJ, Escudero M, Espinosa-Ruiz S, Fay MF, Fernández M, Flanagan NS, Forest F, Fowler RM, Freiberg M, Gallagher RV, Gaya E, Gehrke B, Gelwick K, Grace OM, Granados Mendoza C, Grenié M, Groom QJ, Hackel J, Hagen ER, Hágsater E, Halley J.M, Hu AQ, Jaramillo C, Kattge J, Keith DA, Kirk P, Kissling WD, Knapp S, Kreft H, Kuhnhäuser BG, Larridon I, Leão TCC, Leitch IJ, Liimatainen K, Lim JY, Lucas E, Lücking R, Luján M, Luo A, Magallón S, Maitner B, Márquez-Corro JI, Martín-Bravo S, Martins-Cunha K, Mashau AC, Mauad AV, Maurin O, Medina-Lemos R, Merow C, Michelangeli FA, Mifsud JCO, Mikryukov V, Moat J, Monro AK, Muasya AM, Mueller GM, Muellner-Riehl AN, Nargar K, Negrão R, Nicolson N, Niskanen T, Oliveira-Andrino C, Olmstead RG, Ondo I, Oses L, Parra-Sánchez E, Paton AJ, Pellicer J, Pellissier L, Pennington TD, Pérez-Escobar OA, Phillips C, Pironon S, Possingham H, Prance G, Przelomska NAS, Ramírez-Barahona SA, Renner SS, Rincon M, Rivers MC, Rojas Andrés BM, Romero-Soler KJ, Roque N, Rzedowski J, Sanmartín I, Santamaría-Aguilar D, Schellenberger Costa D, Serpell E, Seyfullah LJ, Shah T, Shen X, Silvestro D, Simpson DA, Šmarda P, Šmerda J, Smidt E, Smith SA, Solano-Gomez R, Sothers C, Soto Gomez M, Spalink D, Sperotto P, Sun M, Suz LM, Svenning JC, Taylor A, Tedersoo L, Tietje M, Trekels M, Tremblay RL, Turner R, Vasconcelos T, Veselý P., Villanueva BS, Villaverde T, Vorontsova MS, Walker BE, Wang Z, Watson M, Weigelt P, Wenk EH, Westrip JRS, Wilkinson T, Willett SD, Wilson KL, Winter M, Wirth C, Wölke FJR, Wright IJ, Zedek F, Zhigila DA, Zimmermann NE, Zuluaga A, Zuntini AR. 2023. State of the world's plants and fungi. United Kingdom: Royal Botanic Gardens, Kew. DOI: https://doi.org/10.34885/wnwn-6s63
- Arida BL, Scopece G, Machado RM, Moraes AP, Forni-Martins E, Pinheiro F. 2021. Reproductive barriers and fertility of two neotropical orchid species and their natural hybrid. *Evolutionary Ecology* 35: 41-64. DOI: <u>https://doi.org/10.1007/s10682-020-10095-5</u>
- Asociación Mexicana de Orquideología. 2005. *Galerías*. <u>http://amo.com.mx/galerias.htm</u> (accessed October 07, 2023).
- Balik HI, Beyhan N. 2020. Xenia and metaxenia affects bioactive compounds of hazelnut. *Turkish Journal of Food* and Agriculture Sciences 2: 42-49. DOI: <u>https://doi.org/10.14744/turkifas.2020.009</u>
- Barreda-Castillo JM, Menchaca-García RA, Morales-Ruiz V. 2023a. Presence of metaxenia in fruits 40 days postpollination in *Vanilla planifolia* Andrews and *V. pompona* Schiede. *Revista Mexicana de Ciencias Agrícolas* 14: 289-293. DOI: <u>https://doi.org/10.29312/remexca.v14i2.2973</u>
- Barreda-Castillo JM, Menchaca-García RA, Pérez-Silva A, Sánchez-Coello NG, Luna-Rodríguez M. 2022. Influence of temperature on the infectivity of *Fusarium oxysporum* f. sp. *vanillae* in *Vanilla planifolia* and in hybrids *V. planifolia* x *V. pompona. Biotecnia* **25**: 177-183. DOI: <u>https://doi.org/10.18633/biotecnia.v25i1.1737</u>
- Barreda-Castillo JM, Monribot-Villanueva JL, Velázquez-Rosas N, Bayman P, Guerrero-Analco JA, Menchaca-García RA. 2023b. Morphological and physio-chemical responses to PEG-induced water stress in *Vanilla planifolia*

and *V. pompona* hybrids. *International Journal of Molecular Sciences* **24**: 4690. DOI: <u>https://doi.org/10.3390/</u> <u>ijms24054690</u>

- Bateman RM. 2020. Implications of next-generation sequencing for the systematics and evolution of the terrestrial orchid genus *Epipactis*, with particular reference to the British Isles. *Kew Bulletin* **75**: 4. DOI: <u>https://doi.org/10.1007/S12225-020-9870-X</u>
- Baulcombe DC, Dean C. 2014. Epigenetic regulation in plant responses to the environment. *Cold Spring Harbor Perspectives in Biology* **6**: a019471. DOI: <u>https://doi.org/10.1101/cshperspect.a019471</u>
- Bernhardt P, Edens-Meier R. 2010. What we think we know vs. what we need to know about orchid pollination and conservation: *Cypripedium* L. as a model lineage. *The Botanical Review* **76**: 204-219. DOI: <u>https://doi.org/10.1007/s12229-010-9042-z</u>
- Bertrand J, Baguette M, Joffard N, Schatz B. 2021. Challenges in the systematics and taxonomy of genera that have recently experienced explosive radiation: the case of orchids of the genus *Ophrys. In*: Grandcolas P, Maurel MC, eds. *Systematics and the Exploration of Life*. United Kingdom: Wiley, pp. 113-134. DOI: <u>https://doi.org/10.1002/9781119476870.ch6</u>
- Bodor P, Gaal M, Toth M. 2008. Metaxenia in apples cv. "Rewena", "Relinda", "Baujade" as influenced by scab resistant pollinizers. *International Journal of Horticultural Science* 14: 11-14. DOI: <u>https://doi.org/10.31421/</u> <u>IJHS/14/3/795</u>
- Bohman B, Flematti GR, Barrow RA, Pichersky E, Peakall R. 2016. Pollination by sexual deception it takes chemistry to work. *Current Opinion in Plant Biology* 32: 37-46. DOI: <u>http://dx.doi.org/10.1016/j.pbi.2016.06.004</u>
- Bonatti PM, Sgarbi E, Del Prete C. 2006. Gynostemium micromorphology and pollination in *Epipactis microphylla* (Orchidaceae). *Journal of Plant Research* **119**: 431-437. DOI: <u>https://doi.org/10.1007/s10265-006-0001-z</u>
- Borba EL, Semir J. 1998. *Bulbophyllum* x *cipoense* (Orchidaceae), a new natural hybrid from the Brazilian "campos rupestres": description and biology. *Lindleyana* 13: 113-120. DOI: <u>http://dx.doi.org/10.2307/3392005</u>
- Borba EL, Shepherd GJ, Semir J. 1999. Reproductive systems and crossing potential in three species of *Bulbophyllum* (Orchidaceae) occurring in Brazilian 'campo rupestre' vegetation. *Plant Systematics and Evolution* 217: 205-214. DOI: <u>https://doi.org/10.1007/BF00984366</u>
- Bory S, Grisoni M, Duval M, Besse P. 2008. Biodiversity and preservation of vanilla: present state of knowledge. *Genetic Resources and Crop Evolution* **55**: 551-557. DOI: <u>https://doi.org/10.1007/s10722-007-9260-3</u>
- Brunschwig C, Rochard S, Pierrat A, Rouger A, Senger-Emonnot P, George G, Raharivelomanana P. 2016. Volatile composition and sensory properties of *Vanilla* × *tahitensis* bring new insights for vanilla quality control. *Journal of the Science Food and Agriculture* **96**: 848-858. DOI: <u>https://doi.org/10.1002/jsfa.7157</u>
- Brzosko E, Mirski P. 2021. Floral nectar chemistry in orchids: A short review and meta-analysis. *Plants* **10**: 2315. DOI: <u>https://doi.org/10.3390/plants10112315</u>
- Buragohain B, Chaturvedi SK, Puro N. 2016. Pollination biology of *Phaius tankervilleae* (Banks ex L'Herit) Bl. (Orchidaceae). *The International Journal of Plant Reproductive Biology* **8**: 75-81.
- Caballero-Villalobos L, Silva-Arias GA, Buzatto CR, Nervo MH, Singer RB. 2017. Generalized food-deceptive pollination in four *Cattleya* (Orchidaceae: Laeliinae) species from Southern Brazil. *Flora* 234: 195-206. DOI: <u>https://</u> doi.org/10.1016/j.flora.2017.07.014_
- Cabrera-Reyes H, Draper D, Marques I. 2021. Pollination in the rainforest: scarce visitors and low effective pollinators limit the fruiting success of tropical orchids. *Insects* 12: 856: DOI: <u>https://doi.org/10.3390/insects12100856</u>
- Calevo J, Bazzicalupo M, Adamo M, Robustelli della Cuna FS, Voyron S, Girlanda M, Duffy KJ, Giovannini A, Cornara L. 2021. Floral trait and mycorrhizal similarity between an endangered orchid and its natural hybrid. *Diversity* 13: 550. DOI: <u>https://doi.org/10.3390/d13110550</u>
- Cantuária PDC, Krahl DRP, Krahl AH, Chiron G, da Silva JBF. 2021. The *Catasetum × sheyllae* (Orchidaceae: Catasetinae), a new natural hybrid from Brazilian Amazon. *Phytotaxa* **527**: 257-265. DOI: <u>https://doi.org/10.11646/phytotaxa.527.4.3</u>
- Capó M, Perelló-Suau S, Rita J. 2022. Preventing inbreeding depression in *Anacamptis coriophora* (Orchidaceae) as a model of food-rewarding orchid. *Plant Ecology* **223**: 423-436. DOI: <u>https://doi.org/10.1007/s11258-022-01221-0</u>

- Castro JB, Perdomo O, Singer RB. 2021. Pollination biology and reproductive success in four Brazilian species of Gomesa (Orchidaceae: Oncidiinae): Specific pollinators, but high pollen loss and low fruit set. Plant Species Biology 37: 132-147. DOI: <u>https://doi.org/10.1111/1442-1984.12361</u>
- Cetzal-Ix W, Carnevali G, Jiménez-Machorro R, Pérez-Silva EA. 2020. Laelia × meavei: A new natural hybrid between L. dawsonii fo. dawsonii and L. rubescens fo. peduncularis (Orchidaceae: Laeliinae) from Oaxaca, Mexico. Phytotaxa 446: 81-94. DOI: <u>https://doi.org/10.11646/phytotaxa.446.2.1</u>
- Chambers A. 2019. Vanilla (Vanilla spp.) breeding. In: Al-Khayri JM, Jain SM, Johnson DV, eds. Advances in Plant Breeding Strategies: Industrial and Food Crops Volume 6. United States of America: Springer, pp. 707-734. DOI: <u>https://doi.org/10.1007/978-3-030-23265-8_18</u>
- Chambers A, Cibrián-Jaramillo A, Karremans AP, Moreno-Martínez D, Hernández-Hernández J, Brym M, Resende MFR, Moloney R, Sierra SN, Hasing T, Alomia YA, Hu Y. 2021. Genotyping-by-sequencing diversity analysis of international Vanilla collections uncovers hidden diversity and enables plant improvement. *Plant Science* 311: 111019. DOI: <u>https://doi.org/10.1016/j.plantsci.2021.111019</u>
- Chandler S, Dunwell JM. 2008. Gene flow, risk assessment and the environmental release of transgenic plants. *Critical Reviews in Plant Sciences* 27: 25-49. DOI: <u>http://dx.doi.org/10.1080/07352680802053916</u>
- Chase MW. 2005. Classification of Orchidaceae in the age of DNA data. *Curtis's Botanical Magazine* 22: 2-7. DOI: https://doi.org/10.1111/j.1355-4905.2005.00466.x
- Chase MW, Cameron K, Freudenstein J, Pridgeon A, Salazar G, van der Berg C, Schuiteman A. 2015. An updated classification of Orchidaceae. *Botanical Journal of the Linnean Society* **177**: 151-174. DOI: <u>https://doi.org/10.1111/boj.12234</u>
- Cheng X, Zhang J, Wang H, Chen T, Li G, Yan C, Jin Q, Lin Y, Cai Y. 2020. Effects of metaxenia on stone cell formation in pear (*Pyrus bretschneideri*) based on transcriptomic analysis and functional characterization of the lignin-related gene PbC4H2. *Forests* **11**: 53. DOI: <u>https://doi.org/10.3390/f11010053</u>
- Chung MY, Nason JD, Chung MG. 2005. Patterns of hybridization and population genetic structure in the terrestrial orchids *Liparis kumokiri* and *Liparis makinoana* (Orchidaceae) in sympatric populations. *Molecular Ecology* 14: 4389-4402. DOI: https://doi.org/10.1111/j.1365-294X.2005.02738.x
- Cozzolino S, Aceto S, Caputo P, Menale B. 1998. Characterization of Orchis x dietrichiana Bogenh., a natural orchid hybrid. Plant Biosystems 132: 71-76. DOI: <u>https://doi.org/10.1080/11263504.1998.10654193</u>
- Cozzolino S, Nardella AM, Impagliazzo S, Widmer A, Lexer C. 2006. Hybridization and conservation of Mediterranean orchids: Should we protect the orchid hybrids or the orchid hybrid zones? *Biological Conservation* 129: 14-23. DOI: <u>https://doi.org/10.1016/j.biocon.2005.09.043</u>
- Cozzolino S, Widmer A. 2005. Orchid diversity: an evolutionary consequence of deception? *Trends in Ecology and Evolution* **20**: 487-494. DOI: <u>https://doi.org/10.1016/j.tree.2005.06.004</u>
- da Silva-Oliveira JP, Garrett R, Bello-Koblitz MG, Furtado-Macedo A. 2022. Vanilla flavor: species from the Atlantic forest as natural alternatives. *Food Chemistry* 375: 131891. DOI: <u>https://doi.org/10.1016/j.foodchem.2021.131891</u>
- Daniell H, Jin S, Zhu X, Gitzendanner MA, Soltis DE, Soltis PS. 2021. Green giant-a tiny chloroplast genome with mighty power to produce high-value proteins: history and phylogeny. *Plant Biotechnology Journal* 19: 430-447. DOI: <u>https://doi.org/10.1111/pbi.13556</u>
- Darwin C. 1862. On the various contrivances by which British and foreign orchids are fertilized by insects, and on the good effect of intercrossing. England: John Murray.
- Dearnaley JDW. 2007. Further advances in orchid mycorrhizal research. *Mycorrhiza* 17: 475-486. DOI: <u>https://doi.org/10.1007/s00572-007-0138-1</u>
- de Jong TJ, Scott RJ. 2007. Parental conflict does not necessarily lead to the evolution of imprinting. *Trends in Plant Science* **12**: 439-443. DOI: <u>https://doi.org/10.1016/j.tplants.2007.07.003</u>
- de Oliveira RT, da Silva-Oliveira JP, Furtado-Macedo A. 2022. Vanilla beyond *Vanilla planifolia* and *Vanilla × tahi-tensis*: taxonomy and historical Notes, reproductive biology, and metabolites. *Plants* **11**: 3311. DOI: <u>https://doi.org/10.3390/plants11233311</u>

- de Paiva-Neto VB, de Aguiar-Torrezan M, Vieira-da Silva MA, Castro-Padilha DR, Constantino-Borel J, Zuffo-Borges MCR. 2022. Self-pollination of the orchid *Cycnoches haagii* from Brazilian Cerrado results in albino phenotype seedlings. *Ornamental Horticulturae* 28: 85-91. DOI: <u>https://doi.org/10.1590/2447-536X.v28i1.2411</u>
- Dellinger AS. 2020. Pollination syndromes in the 21st century: where do we stand and where may we go? *New Phy-tologist* **228**: 1193-1213. DOI: <u>https://doi.org/10.1111/nph.16793</u>
- Deng L, Wang T, Hu J, Yang X, Yao Y, Jin Z, Wang, Z. 2022. Effects of pollen sources on fruit set and fruit characteristics of 'Fengtangli' Plum (*Prunus salicina* Lindl.) based on Microscopic and transcriptomic analysis. *International Journal of Molecular Sciences* 23: 12959. DOI: <u>https://doi.org/10.3390/ijms232112959</u>
- Devadas R, Pattanayak SL, Singh R. 2016. Studies on cross compatibility in *Dendrobium* species and hybrids. *Indian Journal of Genetics and Plant Breeding* **76**: 344-355. DOI: <u>https://doi.org/10.5958/0975-6906.2016.00052.3</u>
- Divakaran M, Babu KN, Peter KV. 2016. Protocols for biotechnological interventions in improvement of Vanilla (Vanilla planifolia Andrews.). In: Jain S, eds. Protocols for In Vitro Cultures and Secondary Metabolite Analysis of Aromatic and Medicinal Plants. Second Edition. United States of America: Humana Press, pp. 47-64. DOI: https://doi.org/10.1007/978-1-4939-3332-7_4
- Divakaran M, Babu KN, Ravindran PN, Peter KV. 2006. Interspecific hybridization in vanilla and molecular characterization of hybrids and selfed progenies using RAPD and AFLP markers. *Scientia Horticulturae* 108: 414-422. DOI: <u>https://doi.org/10.1016/j.scienta.2006.02.018</u>
- dos Santos IS, da Silva MJ. 2023. Anatomy and histochemistry of the vegetative system of *Brachystele guayanensis* (Lindl.) Schltr. (Orchidaceae), a potential medicinal species. *Plants* **12**: 2635. DOI: <u>https://doi.org/10.3390/plants12142635</u>
- Dutra D, Kane ME, Adams CR, Richardson L. 2009. Reproductive biology of *Cyrtopodium punctatum in situ*: implications for conservation of an endangered Florida orchid. *Plant Species Biology* **24**: 92-103. DOI: <u>https://doi.org/10.1111/j.1442-1984.2009.00242.x</u>
- Eckert CG, Herlihy CR. 2004. Using a cost-benefit approach to understand the evolution of self-fertilization in plants: the perplexing case of *Aquilegia canadensis* (Ranunculaceae). *Plant Species Biology* **19**: 159-173. DOI: https://doi.org/10.1111/j.1442-1984.2004.00112.x
- Edens-Meier R, Westhus E, Bernhardt P. 2013. Floral biology of large-flowered *Thelymitra* species (Orchidaceae) and their hybrids in Western Australia. *Journal of Plant Systematics* **15**: 165-183. DOI: <u>https://doi.org/10.7751/</u> telopea2013020
- Edlund AF, Swanson R, Preuss D. 2004. Pollen and stigma structure and function: the role of diversity in pollination. *The Plant Cell* **16**: 84-97. DOI: <u>https://doi.org/10.1105/tpc.015800</u>
- Emeterio-Lara A, García-Franco JG, Hernández-Apolinar M, Mora-Herrera ME, Toledo-Hernández VH, Valencia-Díaz S, Flores-Palacios A. 2018. Endogamy costs and reproductive biology of *Laelia autumnalis*, an endemic orchid of Mexico. *Plant Ecology* 219: 1423-1434. DOI: <u>https://doi.org/10.1007/s11258-018-0891-6</u>
- Endress PK. 2016. Development and evolution of extreme synorganization in angiosperm flowers and diversity: a comparison of Apocynaceae and Orchidaceae. *Annals of Botany* **117**: 749-767. DOI: <u>https://doi.org/10.1093/aob/mcv119</u>
- Evans A, Jacquemyn H. 2020. Impact of mating system on range size and niche breadth in *Epipactis* (Orchidaceae). *Annals of Botany* **126**: 1203-1214. DOI: <u>https://doi.org/10.1093/aob/mcaa142</u>
- Evans A, Jacquemyn H. 2022. Range size and niche breadth as predictors of climate-induced habitat change in *Epipac-tis* (Orchidaceae). *Frontiers in Ecology and Evolution* **10**: 894616. DOI: <u>https://doi.org/10.3389/fevo.2022.894616</u>
- Evans SA, Whigham DF, Hartvig I, McCormick MK. 2023. Hybridization in the fringed orchids: an analysis of species boundaries in the face of gene flow. *Diversity* **15**: 384. DOI: <u>https://doi.org/10.3390/d15030384</u>
- Faegri K, Pijl L. 1979. *The principles of pollination ecology*. Third edition. England: Pergamon Press. ISBN: 0-08-021338-3.
- Fantinato E, Del Vecchio S, Baltieri M, Fabris B, Buffa G. 2017. Are food-deceptive orchid species really functionally specialized for pollinators? *Ecological Research* 32: 951-959. DOI: <u>https://doi.org/10.1007/s11284-017-1501-0</u>

- Favre-Godal Q, Gourguillon L, Lordel-Madeleine S, Gindro K, Choisy P. 2020. Orchids and their mycorrhizal fungi: an insufficiently explored relationship. *Mycorrhiza* **30**: 5-22. DOI: <u>https://doi.org/10.1007/s00572-020-00934-2</u>
- Fay MF. 2018. Orchid conservation: how can we meet the challenges in the twenty-first century? *Botanical Studies* 59: 16. DOI: <u>https://doi.org/10.1186/s40529-018-0232-z</u>
- Fay MF, Smith RJ, Zuiderduin K, Hooper E, Samuel R, Bateman RM, Chase MW. 2007. How does hybridization influence the decision making process in conservation? The genus Orchis (Orchidaceae) as a case history. Lankesteriana 7: 135-137. DOI: <u>https://doi.org/10.15517/lank.v7i1-2.18455</u>
- Ferdy JB, Austerlitz F. 2002. Extinction and introgression in a community of partially cross-fertile plant species. *The American Naturalist* **160**: 74-86. DOI: <u>https://doi.org/10.1086/340606</u>
- Fiorini CF, de Camargo-Smidt E, Knowles LL, Borba EL. 2023. Hybridization boosters diversification in a Neotropical Bulbophyllum (Orchidaceae) group. Molecular Phylogenetics and Evolution 186: 107858. DOI: <u>https://doi.org/10.1016/j.ympev.2023.107858</u>
- Freudenstein JV, Chase MW. 2015. Phylogenetic relationships in Epidendroideae (Orchidaceae), one of the great flowering plant radiations: progressive specialization and diversification. *Annals of Botany* **115**: 665-681. DOI: <u>https://doi.org/10.1093/aob/mcu253</u>
- Gigant RL, De Bruyn A, M'sa T, Viscardi G, Gigord L, Gauvin-Bialecki A, Pailler T, Humeau L, Grisoni M, Besse P. 2016. Combining pollination ecology and fine-scale spatial genetic structure analysis to unravel the reproductive strategy of an insular threatened orchid. *South African Journal of Botany* **105**: 25-35. DOI: <u>http://dx.doi.org/10.1016/j.sajb.2016.02.205</u>
- Givnish TJ, Spalink D, Ames M, Lyon SP, Hunter SJ, Zuluaga A, Iles WJD, Clements MA, Arroyo MTK, Leebens-Mack J, Endara L, Kriebel R, Neubig KM, Whitten WM, Williams NH, Cameron KM. 2015. Orchid phylogenomics and multiple drivers of their extraordinary diversification. *Proceedings of the Royal Society B: Biological Sciences* 282: 20151553. DOI: <u>https://doi.org/10.1098/rspb.2015.1553</u>
- Goulet BE, Roda F, Hopkins R. 2017. Hybridization in plants: old Ideas, new techniques. *Plant Physiology* **173**: 65-78. DOI: <u>https://doi.org/10.1104/pp.16.01340</u>
- Gravendeel B, Smithson A, Silk FJW, Schuiteman A. 2004. Epiphytism and pollinator specialization: drivers for orchid diversity? *Philosophical Transactions of the Royal Society B* 359: 1523-1535. DOI: <u>https://doi.org/10.1098/</u> <u>rstb.2004.1529</u>
- Grisoni M, Nany F. 2021. The beautiful hills: half a century of vanilla (*Vanilla planifolia* Jacks. ex Andrews) breeding in Madagascar. *Genetic Resources and Crop Evolution* **68**: 1691-1708. DOI: <u>https://doi.org/10.1007/s10722-021-01119-2</u>
- Hallett AC, Mitchell RJ, Chamberlain ER, Karron JD. 2017. Pollination success following loss of a frequent pollinator: the role of compensatory visitation by other effective pollinators. *AoB Plants* 9: plx020. DOI: <u>https://doi.org/10.1093/aobpla/plx020</u>
- Havkin-Frenkel D, Belanger FC. 2018. Molecular analysis of a Vanilla hybrid cultivated in Costa Rica. In: Havkin-Frenkel D, Belanger FC, eds. Handbook of Vanilla Science and Technology. Second edition. United States of America: Wiley, pp. 391-402. DOI: <u>https://doi.org/10.1002/9781119377320.ch21</u>
- Hernández-Hernández J. 2018. Mexican Vanilla production. In: Havkin-Frenkel D, Belanger FC, eds. Handbook of Vanilla Science and Technology. Second edition. United States of America: Wiley, pp. 03-26. DOI: <u>https://doi.org/10.1002/9781119377320.ch1</u>
- Hew CS, Yong JWH. 2004. The physiology of tropical orchids in relation to the industry. Second edition. England: World Scientific Publishing. ISBN: 981-238-801-X.
- Hietz P, Wagner K, Nunes Ramos F, Cabral JS, Agudelo C, Benavides AM, Cach-Pérez M.J, Cardelús CL, Chilpa-Galván N, Erickson-Nascimento da Costa L. 2022. Putting vascular epiphytes on the traits map. *Journal of Ecology* 110: 340-358. DOI: <u>https://doi.org/10.1111/1365-2745.13802</u>
- Hinsley A, de Boer H, Fay M, Gale S, Gardiner L, Gunasekara R, Kumar P, Masters S, Metusala D, Roberts L, Veld-

man S, Wong S, Phelps J. 2018. A review of the trade in orchids and its implications for conservation. *Botanical Journal of the Linnean Society* **186**: 435-455. DOI: <u>https://doi.org/10.1093/botlinnean/box083</u>

- Hu C, Jiao Deng X, Tu X, Lu A, Xie C, Jiang K, Zeng X, Liu Z, Huang W, Luo Y. 2022. The ecological adaptation of the unparalleled plastome character evolution in slipper orchids. *Frontiers in Plant Science* 13: 1075098. DOI: <u>https://doi.org/10.3389/fpls.2022.1075098</u>
- Hu A, Gale SW, Liu Z, Suddee S, Hsu T, Fischer GA, Saunders RMK. 2020. Molecular phylogenetics and floral evolution of the *Cirrhopetalum* alliance (*Bulbophyllum*, Orchidaceae): evolutionary transitions and phylogenetic signal variation. *Molecular Phylogenetics and Evolution* 143: 10668. DOI: <u>https://doi.org/10.1016/j.</u> <u>ympev.2019.106689</u>
- Hu Y, Resende MFR, Bombarely A, Brym M, Bassil E, Chambers A. 2019. Genomics-based diversity analysis of Vanilla species using a *Vanilla planifolia* draft genome and genotyping-by-sequencing. *Scientific Reports* 9: 3416. DOI: <u>https://doi.org/10.1038/s41598-019-40144-1</u>
- Humaña AM, Cisternas MA, Valdivia CE. 2008. Breeding system and pollination of selected orchids of the genus *Chloraea* (Orchidaceae) from central Chile. *Flora* **203**: 469-473. DOI: <u>https://doi.org/10.1016/j.flora.2007.08.004</u>
- Hurskainen S, Jakalaniemi A, Kaitala V, Kull T, Motlep M, Ramula S, Tuomi J. 2017. Temporal cycles and spatial asynchrony in the reproduction and growth of a rare nectarless orchid, *Cypripedium calceolus*. *Botanical Journal* of the Linnean Society 183: 316-326. DOI: <u>https://doi.org/10.1093/botlinnean/bow008</u>
- Jersáková J, Johnson SD, Kindlmann P. 2006. Mechanisms and evolution of deceptive pollination in orchids. *Biological Reviews* 81: 219-235. DOI: <u>https://doi.org/10.1017/S1464793105006986</u>
- Jin X, Ren Z, Xu S, Wang H, Li D, Li Z. 2014. The evolution of floral deception in *Epipactis veratrifolia* (Orchidaceae): from indirect defense to pollination. *BMC Plant Biology* 14: 63. DOI: <u>https://doi.org/10.1186/1471-2229-14-63</u>
- Johnson SD. 2000. Batesian mimicry in the non-rewarding orchid *Disa pulchra*, and its consequences for pollinator behaviour. *Biological Journal of the Linnean Society* **71**: 119-132. DOI: https://doi.org/10.1006/bijl.1999.0430
- Johnson SD. 2018. Natural hybridization in the orchid flora of South Africa: comparisons among genera and floristic regions. *South African Journal of Botany* **118**: 290-298. DOI: <u>https://doi.org/10.1016/j.sajb.2018.01.011</u>
- Johnson SD, Edwards TJ. 2000. The structure and function of orchid pollinaria. *In*: Dafni A, Hesse M, Pacini E, eds. *Pollen and pollination*. United States of America: Springer, pp. 243-270. DOI: <u>https://doi.org/10.1007/978-3-7091-6306-1</u>
- Johnson SD, Peter CI, Agren J. 2004. The effect of nectar addition on removal and geitonogamy in the non-rewarding orchid *Anacamptis morio*. *Proceedings of the Royal Society Biological Sciences* **271**:803-809. DOI: <u>https://doi.org/10.1098/rspb.2003.2659</u>
- Johnson TR, Stewart SL, Kauth P, Kane ME, Philman N. 2009. Confronting assumptions about spontaneous autogamy in populations of *Eulophia alta* (Orchidaceae) in south Florida: assessing the effect of pollination treatments on seed formation, seed germination and seedling development. *Botanical Journal of the Linnean Society* 161: 78-88. DOI: https://doi.org/10.1111/j.1095-8339.2009.00992.x
- Jolman D, Batalla MI, Hungerford A, Norwood P, Tait N, Wallace LE. 2022. The challenges of growing orchids from seeds for conservation: An assessment of asymbiotic techniques. *Applications in Plant Sciences* **10**: e11496. DOI: https://doi.org/10.1002/aps3.11496
- Kempe K, Gils M. 2011. Pollination control technologies for hybrid breeding. *Molecular Breeding* 27: 417-437. DOI: <u>https://doi.org/10.1007/s11032-011-9555-0</u>
- Kim Y, Cheon S, Hong J, Kim K. 2023. Evolutionary patterns of the chloroplast genome in vanilloid orchids (Vanilloideae, Orchidaceae). *International Journal of Molecular Sciences* 24: 3808. DOI: <u>https://doi.org/10.3390/jjms24043808</u>
- Kishor R, Sharma GJ. 2009. Intergeneric hybrid of two rare and endangered orchids, *Renanthera imschootiana* Rolfe and *Vanda coerulea* Griff. ex L. (Orchidaceae): synthesis and characterization. *Euphytica* **165**: 247-256. DOI: https://doi.org/10.1007/s10681-008-9755-9_

- Kocyan A, Endress PK. 2001. floral structure and development of *Apostasia* and *Neuwiedia* (apostasioideae) and their relationships to other Orchidaceae. *International Journal of Plant Sciences* **162**: 847-867. DOI: <u>https://doi.org/10.1086/320781</u>
- Kropf M, Renner SS. 2008. Pollinator-mediated selfing in two deceptive orchids and a review of pollinium tracking studies addressing geitonogamy. *Oecologia* 155: 497-508. DOI: <u>https://doi.org/10.1007/s00442-007-0919-4</u>
- Kumar S, Singh A. 2016. Epigenetic regulation of abiotic stress tolerance in plants. *Advances in Plants & Agriculture Research* **5**: 179.
- Kundu A, Karmakar P. 2022. Pollination ecology and breeding system of *Ecolium ligustrinum* (Acanthaceae): a transition from autogamy to xenogamy through specialized plant-pollinator interactions. *Acta Botanica Hungarica* 64: 137-155. DOI: <u>https://doi.org/10.1556/034.64.2022.1-2.7</u>
- Lanzino M, Palermo AM, Pellegrino G. 2023. The effect of inflorescence display size and flower position on pollination success in two deceptive and one rewarding orchid. *Plant Biology* 25: 396-402. DOI: <u>https://doi.org/10.1111/ plb.13505</u>
- Li C, Dong N, Zhao Y, Wu S, Liu Z, Zhai J. 2021a. A review for the breeding of orchids: current achievements and prospects. *Horticultural Plant Journal* 7: 380-392. DOI: <u>https://doi.org/10.1016/j.hpj.2021.02.006</u>
- Li T, Yang W, Wu S, Selosse M, Gao J. 2021b. Progress and prospects of mycorrhizal fungal diversity in orchids. *Frontiers in Plant Science* 12: 646325. DOI: <u>https://doi.org/10.3389/fpls.2021.646325</u>
- Li Y, Ma L, Liu D, Zhao X, Zhang D, Ke S, Chen G, Zheng Q, Liu Z, Lan S. 2023. *Apostasia fujianica* (Apostasioideae, Orchidaceae), a new Chinese species: evidence from morphological, genome size and molecular analyses. *Phytotaxa* 583: 277-284. DOI: <u>https://doi.org/10.11646/phytotaxa.583.3.6</u>
- López-Caamal A, Tovar-Sánchez E. 2014. Genetic, morphological, and chemical patterns of plant hybridization. *Revista Chilena de Historia Natural* 87: 16. DOI: https://doi.org/10.1186/s40693-014-0016-0
- Lozano-Rodríguez MA, Luna-Rodríguez M, Pech-Canché JM, Menchaca-García RA, Cerdán-Cabrera CR. 2022. Visit frequency of Euglossine bees (Hymenoptera: Apidae) to mature fruits of *Vanilla planifolia* (Orchidaceae). *Acta Botanica Mexicana* **129**: e2001. DOI: <u>https://doi.org/10.21829/abm129.2022.2001</u>
- Lu H, Liu Z, Lan S. 2019. Genome sequencing reveals the role of MADS-box gene families in the floral morphology evolution of orchids. *Horticultural Plant Journal* 5: 247-254. DOI: <u>https://doi.org/10.1016/j.hpj.2019.11.005</u>
- Lubinsky P, Cameron KM, Molina MC, Wong M, Lepers-Andrzejewski S, Gómez-Pompa A, Kim S. 2008. Neotropical roots of a Polynesian spice: the hybrid origin of Tahitian vanilla, *Vanilla tahitensis* (Orchidaceae). *American Journal of Botany* **95**: 1040-1047. DOI: <u>https://doi.org/10.3732/ajb.0800067</u>
- Luo H, Xiao H, Liang Y, Liu N, Turner C, Tan C, Xiong D, Yang B. 2021. Batesian mimicry in the nonrewarding saprophytic orchid *Danxiaorchis yangii*. Ecology and Evolution 11: 2524-2534. DOI: <u>https://doi.org/10.1002/ece3.7193</u>
- MacInnis G, Forrest J. 2020. Field design can affect cross-pollination and crop yield in strawberry (*Fragaria x ananassa D.*). Agriculture, Ecosystems and Environment 289: 106738. DOI: <u>https://doi.org/10.1016/j.agee.2019.106738</u>
- Malaviya DR, Roy AK, Kaushal P, Yadav A, Pandey DK. 2019. Complementary gene interaction and xenia effect controls the seed coat colour in interspecific cross between *Trifolium alexandrinum* and *T. apertum. Genetica* 147: 197-203. DOI: <u>https://doi.org/10.1007/s10709-019-00063-5</u>
- Mant J, Peakall R, Weston PH. 2005. Specific pollinator attraction and the diversification of sexually deceptive Chiloglottis (Orchidaceae). *Plant Systematics and Evolution* 253: 185-200. DOI: <u>https://doi.org/10.1007/s00606-005-0308-6</u>
- Marques I, Draper D, Riofrío L, Naranjo C. 2014. Multiple hybridization events, polyploidy and low post mating isolation entangle the evolution of neotropical species of *Epidendrum* (Orchidaceae). *BMC Evolutionary Biology* 14: 20. DOI: <u>https://doi.org/10.1186/1471-2148-14-20</u>
- McCormick MK, Whigham DF, Canchani-Viruet A. 2018. Mycorrhizal fungi affect orchid distribution and population dynamics. *New Phytologist* 219: 1207-1215. DOI: <u>https://doi.org/10.1111/nph.15223</u>
- Menchaca-García RA. 2018. In vitro Propagation of Vanilla. In: Havkin-Frenkel D, Belanger FC, eds. Handbook of Vanilla Science and Technology. Second edition. United States of America: Wiley, pp. 181-190. DOI: <u>https://doi.org/10.1002/9781119377320.ch12</u>

- Menchaca-García RA, Ramos JM, Moreno D, Luna M, Mata M, Vázquez LM, Lozano MA. 2011. *In vitro* germination of *Vanilla planifolia* and *V. pompona* hybrids. *Revista Colombiana de Biotecnología* **13**: 80-84.
- Militaru M, Butac M, Sumedrea D, Chitu E. 2015. Effect of metaxenia on the fruit quality of scab resistant apple varieties. *Agriculture and Agricultural Science Procedia* **6**: 151-156. DOI: <u>https://doi.org/10.1016/j.aaspro.2015.08.052</u>
- Mó E, Ix WC, Carnevali G, Pérez-García E, Basu S. 2014. A new natural hybrid between *Prostechea cochleata* and *P. radiata* (Orchidaceae) from Alta Verapaz, Guatemala. *Turkish Journal of Botany* **38**: 988-998. DOI: <u>https://doi.org/10.3906/bot-1312-86</u>
- Mosquera-Mosquera HR, Valencia-Barrera RM, Acedo C. 2019. Variation and evolutionary transformation of some characters of the pollinarium and pistil in Epidendroideae (Orchidaceae). *Plant Systematics and Evolution* **305**: 353-374. DOI: <u>https://doi.org/10.1007/s00606-019-01575-5</u>
- Naczk A, Kowalkowska AK, Wisniewska N, Halinski LP, Kapusta M, Czerwicka M. 2018. Floral anatomy, ultrastructure and chemical analysis in *Dactylorhiza incarnata/maculata* complex (Orchidaceae). *Botanical Journal of the Linnean Society* 187: 512-536. DOI: <u>https://doi.org/10.1093/botlinnean/boy027</u>
- Ng CKY, Hew CS. 2000. Orchid pseudobulbs- 'false' bulbs with a genuine importance in orchid growth and survival! *Scientia Horticulturae* **83**: 165-172. DOI: <u>https://doi.org/10.1016/S0304-4238(99)00084-9</u>
- Olfati J, Sheykhtaher Z, Qamgosar R, Khasmakhi-Sabet A, Peyvast G, Samizadeh H, Rabiee B. 2010. Xenia and metaxenia on cucumber fruit and seed characteristics. *International Journal of Vegetable Science* **16**: 243-252. DOI: <u>https://doi.org/10.1080/19315260903584167</u>
- Ospina-Calderón NH, Duque-buitrago CA, Tremblar RL, Otero JT. 2015. Pollination ecology of *Rodriguezia* granadensis (Orchidaceae). Lankesteriana 15: 129-139. DOI: <u>https://doi.org/10.15517/lank.v15i2.20745</u>
- Otero JT, Ackerman JD, Bayman P. 2004. Differences in mycorrhizal preferences between two tropical orchids. *Molecular Ecology* **13**: 2393-2404. DOI: <u>https://doi.org/10.1111/j.1365-294X.2004.02223.x</u>
- Otero JT, Flanagan NS, Herre EA, Ackerman JD, Bayman P. 2007. Widespread mycorrhizal specificity correlates to mycorrhizal function in the neotropical, epiphytic orchid *Ionopsis utricularioides* (Orchidaceae). *American Journal of Botany* **94**: 1944-1950. DOI: <u>https://doi.org/10.3732/ajb.94.12.1944</u>
- Pang C, Saunders RM. 2015. Floral biology and pollination ecology of *Desmos chinensis* (Annonaceae): assessing the efficacy of floral synchrony for promoting xenogamy. *International Journal of Plant Sciences* 176: 333-345. DOI: <u>https://doi.org/10.1086/680402</u>
- Pansarin ER. 2003. Biologia floral de *Cleistes macrantha* (Barb. Rodr.) Schltr. (Orchidaceae: Vanilloideae: Pogoniinae). *Brazilian Journal of Botany* **26**: 73-80. DOI: <u>https://doi.org/10.1590/S0100-84042003000100008</u>
- Pansarin ER. 2016. Recent advances on evolution of pollination systems and reproductive biology of vanilloideae (Orchidaceae). *Lankesteriana* **16**: 255-267. DOI: <u>http://dx.doi.org/10.15517/lank.v16i2.26010</u>
- Pansarin ER, Ferreira AWC. 2015. Butterfly pollination in Pteroglossa (Orchidaceae, Orchidoideae): a comparative study on the reproductive biology of two species of a Neotropical genus of Spiranthinae. *Journal of Plant Research* 128: 459-468. DOI: <u>https://doi.org/10.1007/s10265-015-0707-x</u>
- Pansarin ER, Ferreira AWC. 2021. Evolutionary disruption in the pollination system of *Vanilla* (Orchidaceae). *Plant Biology* 24: 157-167. DOI: <u>https://doi.org/10.1111/plb.13356</u>
- Pansarin ER, Pansarin LM. 2016. Crane flies and microlepidoptera also function as pollinators in Epidendrum (Orchidaceae: Laeliinae): the reproductive biology of *E. avicula*. *Plant Species Biology* **32**: 200-209. DOI: <u>https://doi.org/10.1111/1442-1984.12140</u>
- Pansarin LM, Pansarin ER, Sazima M. 2008. Reproductive biology of *Cyrtopodium polyphyllum* (Orchidaceae): a Cyrtopodiinae pollinated by deceit. *Plant Biology* **10**: 650-659. DOI: <u>https://doi.org/10.1111/j.1438-8677.2008.00060.x</u>
- Park HS, Lee WK, Lee SC, Lee HO, Joh HJ, Park JY, Yang TJ. 2021. Inheritance of chloroplast and mitochondrial genomes in cucumber revealed by four reciprocal F1 hybrid combinations. *Scientific Reports* **11**: 2506. DOI: https://doi.org/10.1038/s41598-021-81988-w
- Paun O, Bateman RM, Fay MF, Luna JA, Moat J, Hedrén M, Chase MW. 2011. Altered gene expression and eco-

logical divergence in sibling allopolyploids of *Dactylorhiza* (Orchidaceae). *Evolutionary Biology* **11**: 113. DOI: <u>https://doi.org/10.1186/1471-2148-11-113</u>

- Pavarese G, Trnachida-Lombardo V, Galesi R, D'Emerico S, Casotti R, Cristaudo A, Cozzolino S. 2013. When polyploidy and hybridization produce a fuzzy taxon: the complex origin of the insular neoendemic *Neotinea commuta*ta (Orchidaceae). *Botanical Journal of the Linnean Society* **173**: 707-720. DOI: https://doi.org/10.1111/boj.12093
- Pedersen HA, Ehlers BK. 2000. Local evolution of obligate autogamy in *Epipactis helleborine* subsp. *neerlandica* (Orchidaceae). *Plant Systematics and Evolution* **223**: 173-183. DOI: <u>https://doi.org/10.1007/BF00985277</u>
- Pedersen HA, Srimuang K, Bänziger H, Watthana S. 2018. Pollination-system diversity in Epipactis (Orchidaceae): new insights from studies of *E. flava* in Thailand. *Plant Systematics and Evolution* **304**: 895-909. DOI: <u>https://doi.org/10.1007/s00606-018-1520-5</u>
- Pellegrino G, Bellusci F, Palermo AM. 2017. Functional differentiation in pollination processes among floral traits in *Serapias* species (Orchidaceae). *Ecology and Evolution* 7: 7171-7177. DOI: <u>https://doi.org/10.1002/ece3.3264</u>
- Pemberton RW. 2013. Pollination of Slipper Orchids (Cypripedioideae): a Review. *Lankesteriana* 13: 65-73. DOI: https://doi.org/10.15517/lank.v0i0.11539
- Pérez-Silva A, Nicolás-García M, Petit T, Dijoux J, Vivar-Vera M, Besse P, Grisoni M. 2021. Quantification of the aromatic potential of ripe fruit of *Vanilla planifolia* (Orchidaceae) and several of its closely and distantly related species and hybrids. *European Food Research and Technology* 247: 1489-1499. DOI: <u>https://doi.org/10.1007/s00217-021-03726-w</u>
- Phelps J. 2015. A blooming trade Illegal trade of ornamental orchids in Mainland Southeast Asia (Thailand, Lao PDR, Myanmar). Malaysia: TRAFFIC. ISBN: 978-983-3393-46-6
- Phillips RD, Reiter N, Peakall R. 2020. Orchid conservation: from theory to practice. *Annals of Botany* **126**: 345-362. DOI: <u>https://doi.org/10.1093/aob/mcaa093</u>
- Pinheiro F, de Barros F, Palma-Silva C, Meyer D, Fay MF, Suzuki RM, Lexer C, Cozzolino S. 2010. Hybridization and introgression across different ploidy levels in the Neotropical orchids *Epidendrum fulgens* and *E. puniceoluteum* (Orchidaceae). *Molecular Ecology* 19: 3981-3994. DOI: <u>https://doi.org/10.1111/j.1365-294X.2010.04780.x</u>
- Piotto F, Batagin-Piotto K, de Almeida M, Oliveira G. 2013. Interspecific xenia and metaxenia in seeds and fruits of tomato. *Scientia Agricola* 70: 102-107. DOI: <u>https://doi.org/10.1590/S0103-90162013000200007</u>
- Porras-Alfaro A, Bayman P. 2007. Mycorrhizal fungi of *Vanilla*: diversity, specificity and effects on seed germination and plant growth. *Mycologia* 99: 510-525. DOI: <u>https://doi.org/10.1080/15572536.2007.11832545</u>
- Pramanik D, Spaans M, Kranenburg T, Bogarin D, Heijungs R, Lens F, Smets E, Gravendeel B. 2022. Inflorescence lignification of natural species and horticultural hybrids of Phalaenopsis orchids. *Scientia Horticulturae* 295: 110845. DOI: <u>https://doi.org/10.1016/j.scienta.2021.110845</u>
- Preston CD, Pearman DA. 2015. Plant hybrids in the wild: evidence from biological recording. *Biological Journal* of the Linnean Society **115**: 555-572. DOI: <u>https://doi.org/10.1111/bij.12577</u>
- Pridgeon AM. 2014. Introduction. *In*: Gregory M, Cutler DF, eds. *Anatomy of the monocotyledons X. Orchidaceae*. England: Oxford University Press. ISBN: 978-0-19-968907-1.
- Radak BD, Vlku AZ, Peskanov JM, Matevski VS, Anackov GT. 2019. Morphological characterization of three natural hybrid orchid taxa, new for Serbia, Montenegro and North Macedonia. *Archives of Biological Sciences* **71**: 596-607. DOI: <u>https://doi.org/10.2298/ABS190520042R</u>
- Ramya M, Jang S, An H, Lee S, Park P, Park PH. 2020. Volatile organic compounds from orchids: from synthesis and function to gene regulation. *International Journal of Molecular Sciences* 21: 1160. DOI: <u>https://doi.org/10.3390/</u> <u>ijms21031160</u>
- Rasmussen HN. 2002. Recent developments in the study of orchid mycorrhiza. *Plant and Soil* 244: 149-163. DOI: https://doi.org/10.1023/A:1020246715436
- Rasmussen HN, Rasmussen FN. 2018. The epiphytic habitat on a living host: reflections on the orchid–tree relationship. *Botanical Journal of the Linnean Society* 186: 456-472 DOI: <u>https://doi.org/10.1093/botlinnean/box085</u>
- Ray HA, Gillett-Kaufman JL. 2022. By land and by tree: pollinator taxa diversity of terrestrial and epiphytic orchids. *Journal of Pollination Ecology* 32: 174-185. DOI: <u>https://doi.org/10.26786/1920-7603(2022)671</u>

- Sabir A. 2014. Xenia and metaxenia in grapes: differences in berry and seed characteristics of maternal grape cv. 'Narince' (*Vitis vinifera* L.) as influenced by different pollen sources. *Plant Biology* 17: 567-573. DOI: <u>https://doi.org/10.1111/plb.12266</u>
- Salazar GA, Jiménez-Machorro R, Huerta HM, Hágsater E. 2014. A new species and a new natural hybrid of Laelia (Orchidaceae) from Oaxaca, Mexico. *Phytotaxa* **178**: 161-170. DOI: <u>http://dx.doi.org/10.11646/phytotaxa.178.3.1</u>
- Sao Leao LC, de Sá-Haiad B, Araujo-Rodarte AT, Pimentel RR, Rodriges-Benevides C, de Santiago-Fernandes LDR, Alves de Lima H. 2019. Reproductive biology of two synchronopatric neotropical species of *Epidendrum* (Orchidaceae). *Flora* 251: 95-104. DOI: <u>https://doi.org/10.1016/j.flora.2019.01.003</u>
- Sari H, Eker T, Sari D, Aksoy M, Bakir M, Dogdu V, Toker C, Canci H. 2023. The Fastest and Most Reliable Identification of True Hybrids in the Genus *Pisum L. Life* 13: 2222. DOI: <u>https://doi.org/10.3390/life13112222</u>
- Sarsaiya S, Shi S, Chen J. 2019. A comprehensive review on fungal endophytes and its dynamics on Orchidaceae plants: current research, challenges, and future possibilities. *Bioengineered* **10**: 316-334. DOI: <u>https://doi.org/10.1080/21655979.2019.1644854</u>
- Sasikumar B. 2010. Vanilla breeding-a review. Agricultural Reviews 31 (2): 139-144.
- Sattler, M, Carvalho, C, Clarindo, W. 2016. The polyploidy and its key role in plant breeding. *Planta* 243: 281-296. DOI: <u>https://doi.org/10.1007/s00425-015-2450-x</u>
- Scopece G, Cozzolino S, Dafni A. 2014. Darwin's orchids of the English and Eurasian countrysides. *In*: Edens-Meier R, Bernhardt P, eds. *Darwin's Orchids Then and Now*. United States of America: The University of Chicago Press, pp. 23-70. ISBN: 978-0-226-17364-1
- Selosse M, Petrolli R, Mujica MI, Laurent L, Perez-Lamarque B, Figura T, Bourceret A, Jacquemyn H, Li T, Gao J, Minasiewicz J, Martos F. 2022. The waiting room hypothesis revisited by orchids: were orchid mycorrhizal fungi recruited among root endophytes? *Annals of Botany* 129: 259-270. DOI: <u>https://doi.org/10.1093/aob/mcab134</u>
- Shahsavar A, Shahhoseini A. 2022. The metaxenia effects of different pollen grains on secondary metabolites enzymes and sugars of 'Piarom' date palm fruit. *Scientific Reports* **12**: 10058. DOI: <u>https://doi.org/10.1038/s41598-022-14373-w</u>
- Shefferson RP, Jacquemyn H, Kull T, Hutchings MJ. 2020 The demography of terrestrial orchids: life history, population dynamics and conservation. *Botanical Journal of the Linnean Society* **192**: 315-332. DOI: <u>https://doi.org/10.1093/botlinnean/boz084</u>
- Shrestha M, Dyer AG, Dorin A, Ren ZX, Burd M. 2020. Rewardlessness in orchids: how frequent and how rewardless? *Plant Biology* 22: 555-561. DOI: <u>https://doi.org/10.1111/plb.13113</u>
- Sletvold N, Grindeland JM, Zu P, Agren J. 2012. Strong inbreeding depression and local outbreeding depression in the rewarding orchid *Gymnadenia conopsea*. *Conservation Genetics* **13**: 1305-1315. DOI: <u>https://doi.org/10.1007/s10592-012-0373-7</u>
- Soltis PS, Folk RA, Soltis DE. 2019. Darwin review: angiosperm phylogeny and evolutionary radiations. *Proceedings The Royal Society Publishing* **286**: 20190099. DOI: <u>http://dx.doi.org/10.1098/rspb.2019.0099</u>
- Squirrell J, Hollingsworth PM, Bateman RM, Tebbitt MC, Hollingsworth ML. 2002. Taxonomic complexity and breeding system transitions: conservation genetics of the *Epipactis leptochila* complex (Orchidaceae). *Molecular Ecology* 11: 1957-1964. DOI: <u>https://doi.org/10.1046/j.1365-294X.2002.01610.x</u>
- Sramkó G, Paun O, Brandrud MK, Laczko L, Molnár A, Bateman RM. 2019. Iterative allogamy-autogamy transitions drive actual and incipient speciation during the ongoing evolutionary radiation within the orchid genus *Epipactis* (Orchidaceae). *Annals of Botany* 124: 481-497. DOI: <u>https://doi.org/10.1093/aob/mcz103</u>
- Srimuang K, Watthana S, Pedersen HA, Rangsayatorn N, Eungwanichayapant PD. 2010. Flowering phenology, floral display and reproductive success in the Genus *Sirindhornia* (Orchidaceae): a comparative study of three pollinator-rewarding species. *BioOne* 47: 439-448. DOI: <u>https://doi.org/10.5735/085.047.0603</u>
- Stebbins G. 1958. The inviability, weakness, and sterility of interspecific hybrids. *Advances in Genetics* **9**: 147-215. https://doi.org/10.1016/S0065-2660(08)60162-5
- Stípková Z, Tsiftsis S, Kindlmann P. 2020. Pollination mechanisms are driving orchid distribution in space. *Scientific Reports* 10: 850. DOI: <u>https://doi.org/10.1038/s41598-020-57871-5</u>

- Stull GW, Pham KK, Soltis PS, Soltis DE. 2023. Deep reticulation: the long legacy of hybridization in vascular plant evolution. *The Plant Journal* 114: 743-766. DOI: <u>https://doi.org/10.1111/tpj.16142</u>
- Suaib S, Mamma S, Rakian T, Suleman D. 2020. Xenia and metaxenia in maize hybrid varieties as a consequence of paternal pollen effect. *Journal of Agronomy* 19: 24-30. DOI: <u>https://doi.org/10.3923/ja.2020.24.30</u>
- Suetsugu K. 2015. Autonomous self-pollination and insect visitors in partially and fully mycoheterotrophic species of *Cymbidium* (Orchidaceae). *Journal of Plant Research* 128: 115-125. DOI: <u>https://doi.org/10.1007/s10265-014-0669-4</u>
- Suetsugu K. 2020. Gynomonoecy in a mycoheterotrophic orchid *Eulophia zollingeri* with autonomous selfing hermaphroditic flowers and putatively outcrossing female flowers. *PeerJ* 8: e10272. DOI: <u>https://doi.org/10.7717/</u> peerj.10272
- Suetsugu K, Fukushima S. 2014. Pollination biology of the endangered orchid *Cypripedium japonicum* in a fragmented forest of Japan. *Plant Species Biology* 29: 294-299. DOI: <u>https://doi.org/10.1111/1442-1984.12016</u>
- Sugiura N. 2013. Pollination and floral ecology of *Arundina graminifolia* (Orchidaceae) at the northern border of the species' natural distribution. *Journal of Plant Research* **127**: 131-139. DOI: <u>https://doi.org/10.1007/s10265-013-0587-x</u>
- Swingle W. 1928. Metaxenia in the date palm, possibly a hormone action by the embryo or endosperm. *Journal of Heredity* **19**: 257-268. DOI: <u>https://doi.org/10.1093/oxfordjournals.jhered.a102996</u>
- Talalaj I, Brzosko E. 2008. Selfing potential in *Epipactis palustris*, *E. helleborine* and *E. atrorubens* (Orchidaceae). *Plant Systematics and Evolution* **276**: 21-29. DOI: <u>https://doi.org/10.1007/s00606-008-0082-3</u>
- Talalaj I, Ostrowiecka B, Wlostowska E, Rutkowska A, Brzosko E. 2017. The ability of spontaneous autogamy in four orchid species: *Cephalanthera rubra*, *Neottia ovata*, *Gymnadenia conopsea*, and *Platanthera bifolia*. *Acta Biologica Cracoviensia* 59: 51-61. DOI: <u>https://doi.org/10.1515/abcsb-2017-0006</u>
- Talalaj I, Skiercynski M. 2015. Mechanism of spontaneous autogamy in the allogamous lepidopteran orchid *Gymnadenia conopsea* (l.) r. br. (Orchidaceae). Acta Biologica Cracoviensia 57: 130-140. DOI: <u>https://doi.org/10.1515/abcsb-2015-0010</u>
- Tatsuzawa F, Saito N, Seki H, Yokoi M, Yukawa T, Shinoda K, Honda T. 2004. Acylated anthocyanins in the flowers of *Vanda* (Orchidaceae). *Biochemical Systematics and Ecology* **32**: 651-664. DOI: <u>https://doi.org/10.1016/j.</u> <u>bse.2004.02.004</u>
- IUCN [The IUCN red list of threatened species]. 2023. Orchidaceae-family. <u>https://www.iucnredlist.org/</u> (accessed September 29, 2023).
- The Royal Horticultural Society. 2023. Orchid hybrid lists. <u>https://www.rhs.org.uk/plants/plantsmanship/plant-regis-tration/orchid-hybrids/orchid-hybrid-lists</u> (accessed October 03, 2023).
- Tremblay RL, Ackerman JD, Zimmerman JK, Calvo RN. 2005. Variation in sexual reproduction in orchids and its evolutionary consequences: a spasmodic journey to diversification. *Biological Journal of Linnean Society* 84: 1-54. DOI: <u>https://doi.org/10.1111/j.1095-8312.2004.00400.x</u>
- Turchetto C, Segatto AL, Turchetto-Zolet AC. 2022. Biotic and abiotic factors in promoting the starting point of hybridization in the Neotropical flora: implications for conservation in a changing world. *Botanical Journal of the Linnean Society* 200: 285-302. DOI: <u>https://doi.org/10.1093/botlinnean/boac042</u>
- Valdivia CE, Cisternas MA, Verdugo GS. 2010. Reproductive biology aspects of two species of the genus *Gavilea* (Orchidaceae, Chloraeinae) in populations from Central Chile. *Gayana Botanica* **67**: 44-51. DOI: <u>https://doi.org/10.4067/S0717-66432010000100005</u>
- Vale A, Rojas D, Álvarez JC, Navarro L. 2010. Breeding system and factors limiting fruit production in the nectarless orchid *Broughtonia lindenii*. *Plant Biology* 13: 51-61. DOI: <u>https://doi.org/10.1111/j.1438-8677.2010.00366.x</u>
- van Esse H, Reuber T, van der Does D. 2020. Genetic modification to improve disease resistance in crops. New Phytologist 225: 70-86. DOI: <u>https://doi.org/10.1111/nph.15967</u>
- Varela-Quirós E. 2010. Vanilla Production in Costa Rica. In: Havkin-Frenkel D, Belanger FC, eds. Handbook of Vanilla Science and Technology. Second edition. United States of America: Wiley, pp. 41-52. DOI: <u>https://doi.org/10.1002/9781444329353.ch3</u>

- Vereecken NJ, Dafni A, Cozzolino S. 2010. Pollination syndromes in Mediterranean orchids-implications for speciation, taxonomy and conservation. *The Botanical Review* 76: 220-240. DOI: <u>https://doi.org/10.1007/s12229-010-9049-5</u>
- Vij SP, Pathak P. 2012. Orchid Diversity: Conservation and utilization. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences 82: 295-300. DOI: <u>https://doi.org/10.1007/s40011-012-0107-0</u>
- Vilcherrez-Atoche JA, Liyama CM, Cardoso JC. 2022. Polyploidization in orchids: from cellular changes to breeding applications. *Plants* **11**: 469. DOI: <u>https://doi.org/10.3390/plants11040469</u>
- Watteyn C, Reubens B, Azofeita-Bolaños JB, Solano-Campos F, Pérez-Silva A, Karremans AP, Muys B. 2023. Cultivation potential of Vanilla crop wild relatives in two contrasting land use systems. *European Journal of Agronomy* 149: 126890. DOI: <u>https://doi.org/10.1016/j.eja.2023.126890</u>
- Willmer P. 2011. Pollination and floral ecology. England: Princeton University Press. ISBN: 978-0-691-12861-0.
- Wu Z, Li W, Chen J, Zhai J, Xu H, Ni L, Wu S. 2019. Chemical constituents and biological activity profiles on *Pleione* (Orchidaceae). *Molecules* 24: 3195. DOI: <u>https://doi.org/10.3390/molecules24173195</u>
- Wurz A, Grass I, Tscharntke T. 2021. Hand pollination of global crops-a systematic review. Basic and Applied Ecology 56: 299-321. DOI: <u>https://doi.org/10.1016/j.baae.2021.08.008</u>
- Yeh C, Chen K, Lee Y. 2021. Asymbiotic germination of *Vanilla planifolia* in relation to the timing of seed collection and seed pretreatments. *Botanical Studies* 62: 6. DOI: <u>https://doi.org/10.1186/s40529-021-00311-y</u>
- Yeung E. 2017. A perspective on orchid seed and protocorm development. *Botanical Studies* **58**: 33. DOI: <u>https://doi.org/10.1186/s40529-017-0188-4</u>
- Yin Y, Zhong P, Zhang G, Chen L, Zeng S, Li M, Liu Z. 2016. Morphological, genome-size and molecular analyses of *Apostasia fogangica* (Apostasioideae, Orchidaceae), a new species from China. *Phytotaxa* 277: 059-067. DOI: https://doi.org/10.11646/phytotaxa.277.1.5
- Zhang Z, Gale SW, Li JH, Fisher GA, Ren MX, Song XQ. 2019. Pollen-mediated gene flow ensures connectivity among spatially discrete sub-populations of *Phalaenopsis pulcherrima*, a tropical food-deceptive orchid. *BMC Plant Biology* **19**: 597. DOI: <u>https://doi.org/10.1186/s12870-019-2179-y</u>
- Zhang X, Jia Y, Chen D, Luo Y, Niu S. 2021. Challenges and perspectives in the study of self-incompatibility in orchids. *International Journal of Molecular Sciences* **22**: 12901. DOI: <u>https://doi.org/10.3390/ijms222312901</u>
- Zhang S, Yang Y, Li J, Qin J, Zhang W, Huang W, Hu H. 2018. Physiological diversity of orchids. *Plant Diversity* **40**: 196-208. DOI: <u>https://doi.org/10.1016/j.pld.2018.06.003</u>
- Zotz G, Winkler U. 2013. Aerial roots of epiphytic orchids: the velamen radicum and its role in water and nutrient uptake. *Oecologia* **171**: 733-741. DOI: <u>https://doi.org/10.1007/s00442-012-2575-6</u>

Associate editor: Arturo de Nova

Author contributions: JMBC Conceptualization, Formal analysis, Writing, ERP Supervision, Writing, JLMV Supervision, Writing, JAGA Funding acquisition, Supervision, Writing, RAMG Funding acquisition, Supervision, Writing.

Supporting Agencies: This research was funded by CONAHCYT (Consejo Nacional de Humanidades, Ciencia y Tecnología) grant to JMBC (CVU: 960142), and research funding assigned to RAMG and JAGA from CITRO and INECOL, respectively.

Conflict of interest: No conflict of interests The authors declare that there is no conflict of interest, financial or personal, in the information, presentation of data and results of this article.